

The CyTOF

Practical considerations for successful
data acquisition

Matthew Cochran, MS

University of Rochester Flow Cytometry Resource

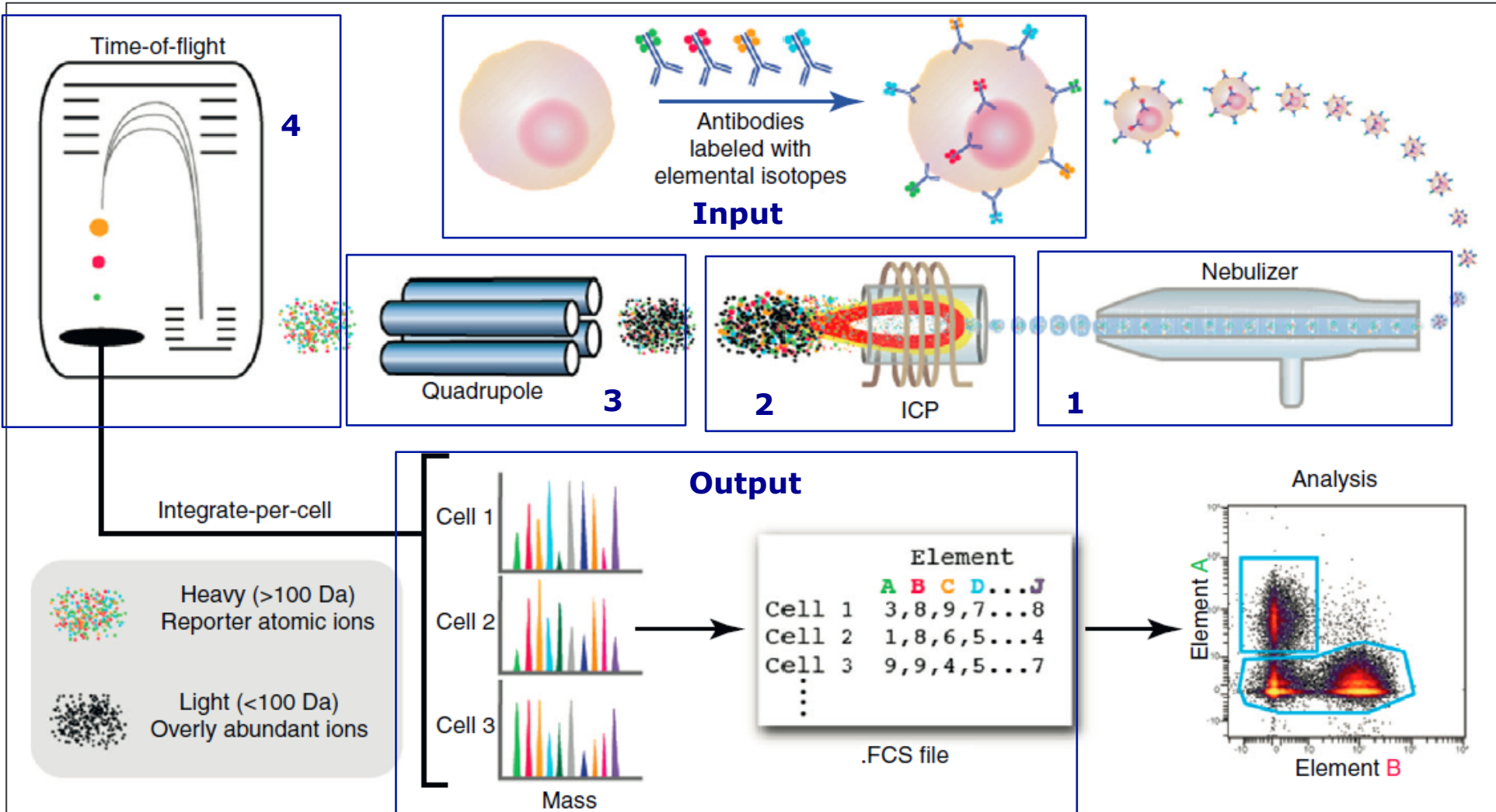
matthew_cochran@urmc.rochester.edu

MEDICINE *of* THE HIGHEST ORDER



**Some slides adapted directly from Fluidigm/DVS Biosciences

Summarizing the System



Bendall S. *et al.* (2012). Trends in Immunology, **33** (7), 323-332

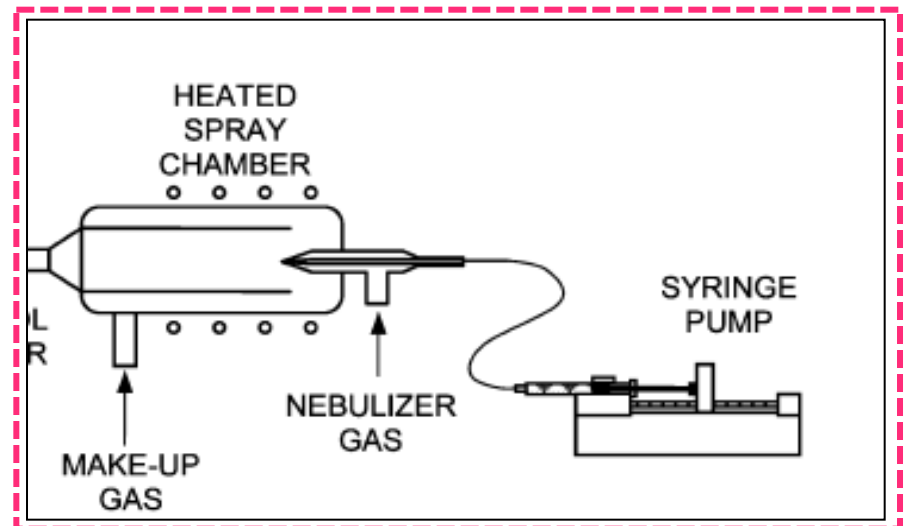
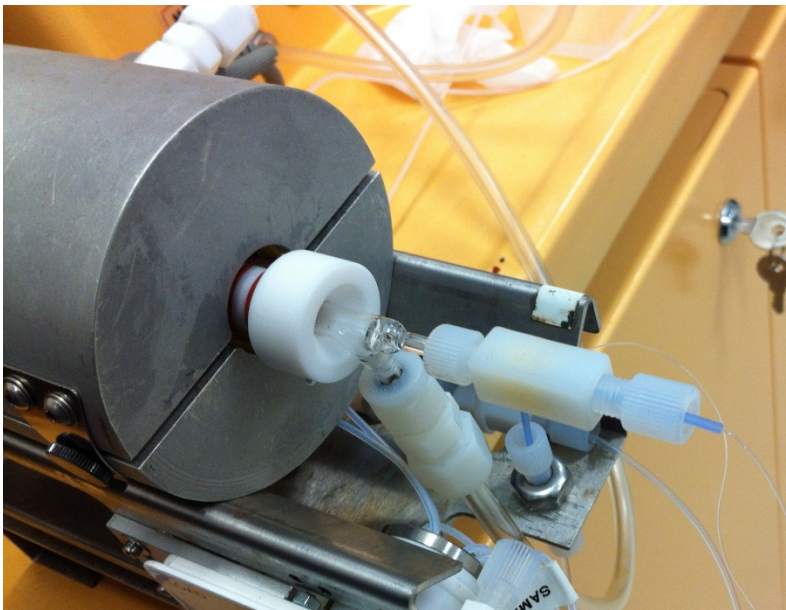
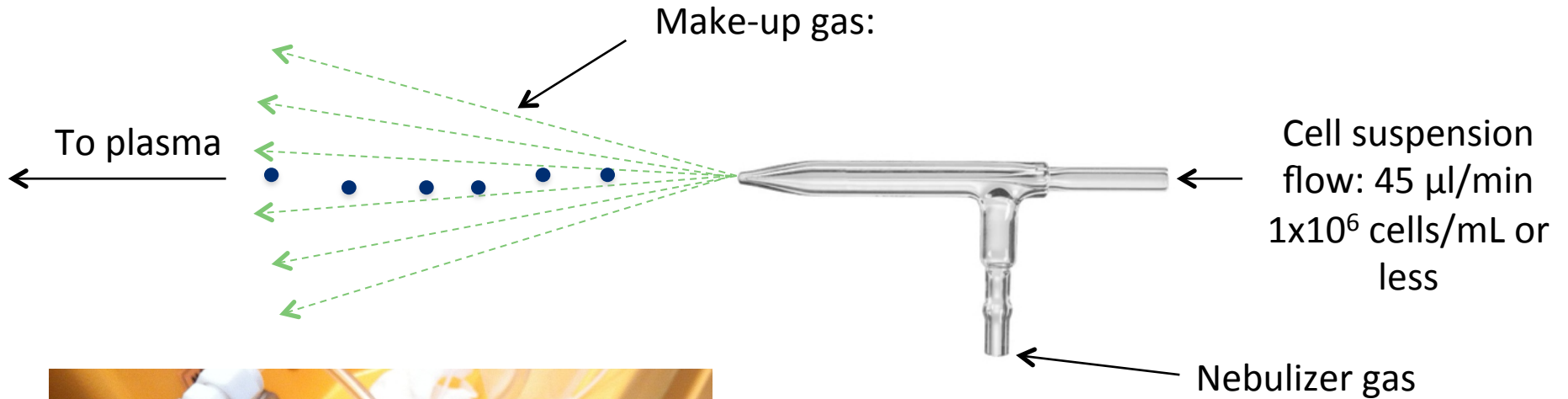
TRENDS in Immunology

Input: Sample Introduction

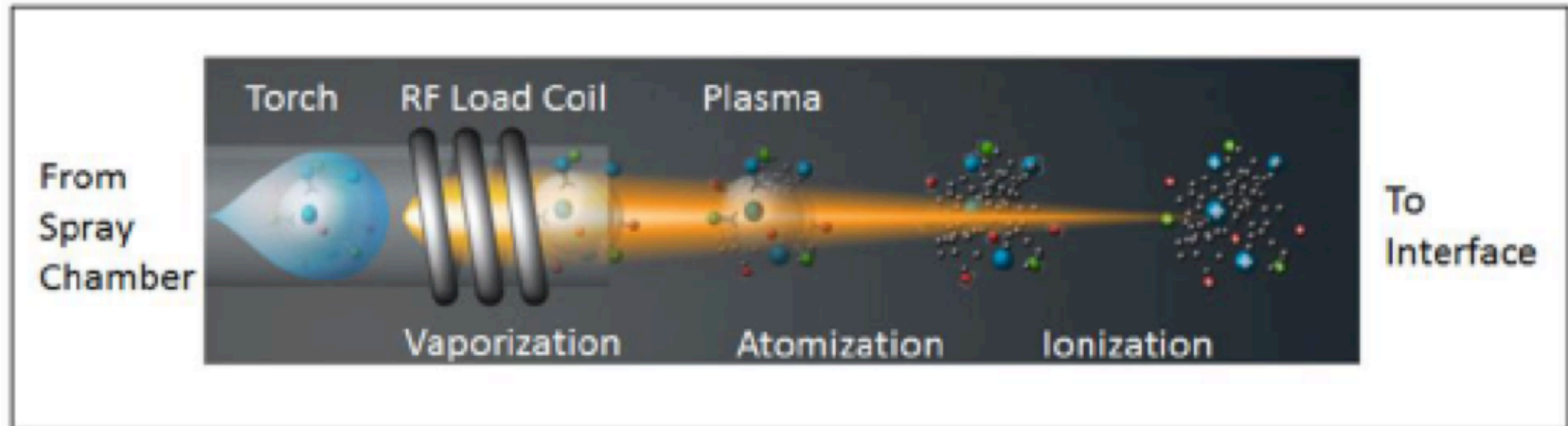
Sample concentration can effect data quality

- Maximum ~500 events per second during run (CyTOF 1-2).
 - The rate that event ion clouds can be collected.
 - Sample introduction at .045 mL/min (used to be .06 mL/min)
 - Not all events are cells.
- Maximum 5 pushes/hr – each push = 10 minutes + washing
 - 4-5 samples with one push each
 - 2-3 samples with 2 pushes
- Maximum $\sim 2.5 \times 10^6$ cells run per hour

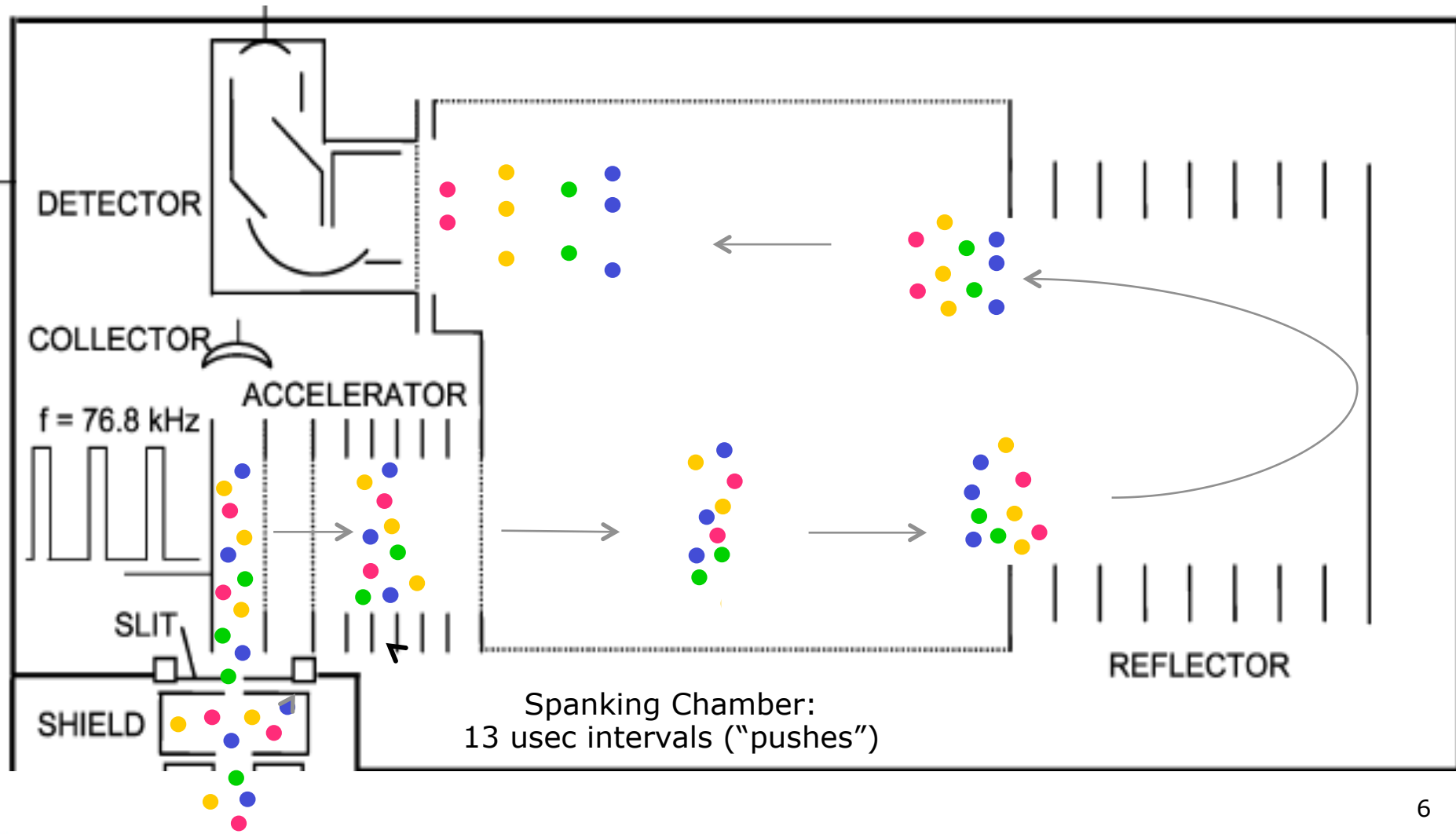
1. Sample Introduction



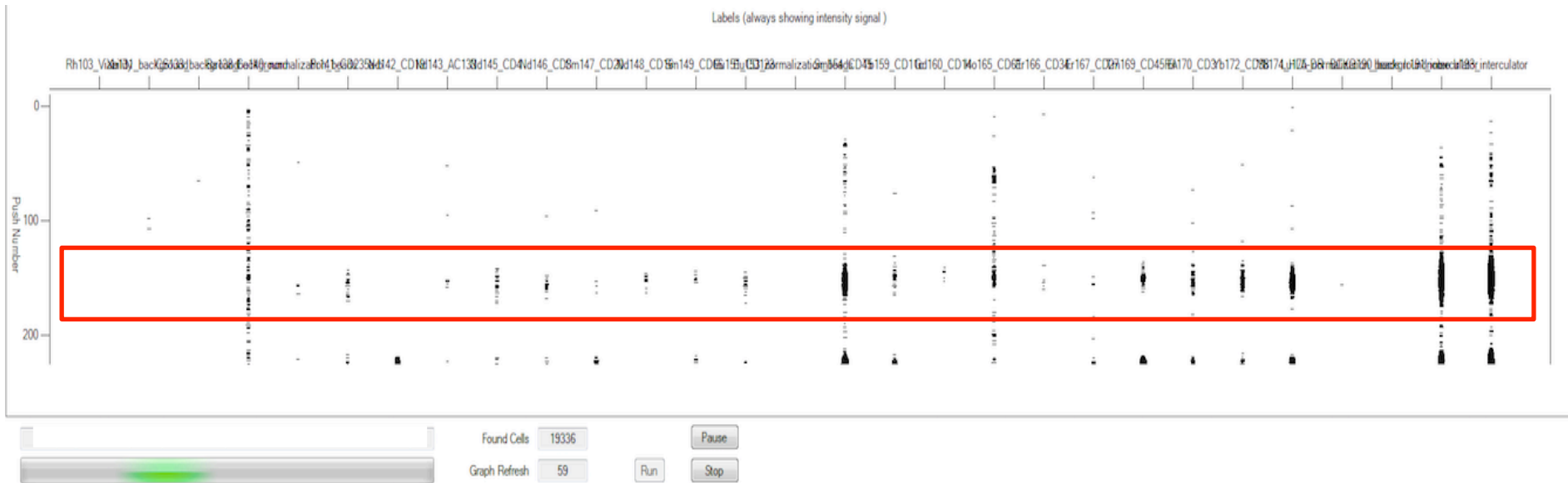
2. Ionization



4. The TOF Chamber



The Raindrop Display



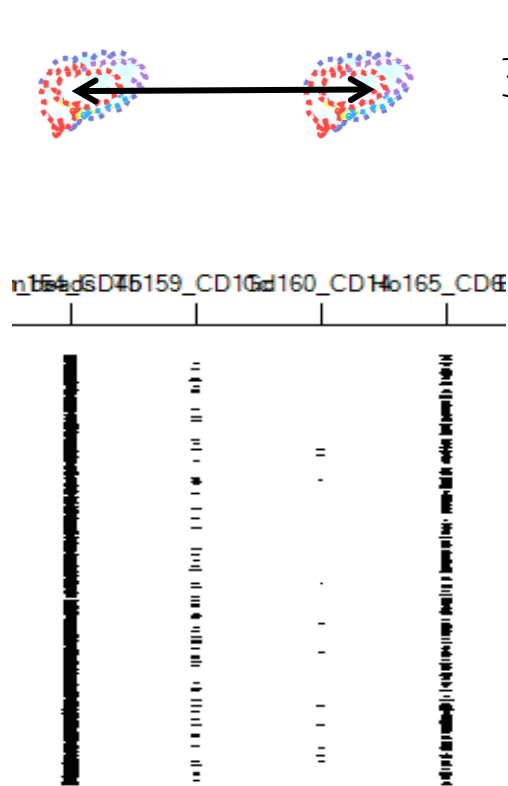
- 29 Parameters Total
- 200 pushes shown
- Some background in the Ba channel
- Good event rate ~ 300 evt/sec
- Software processes the raw data to generate the FCS file.
 - System also generates a "raw" IMD file

Sample Concentration and Cleanliness

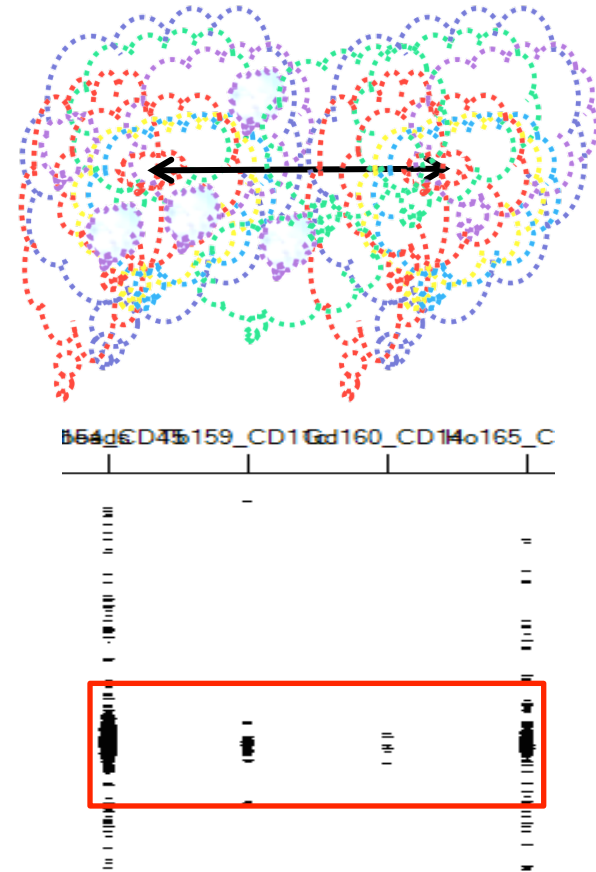
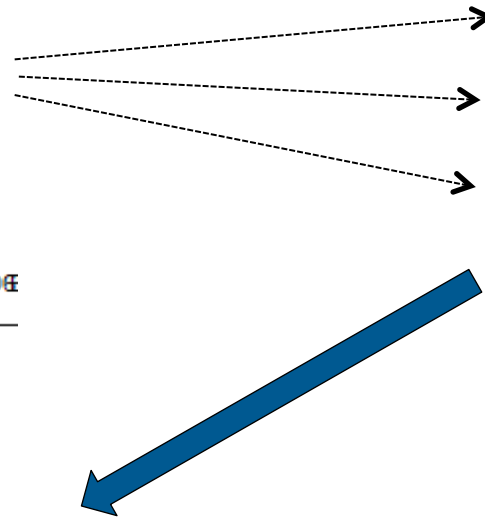
Too close

Diffusion

Cloud Fusion

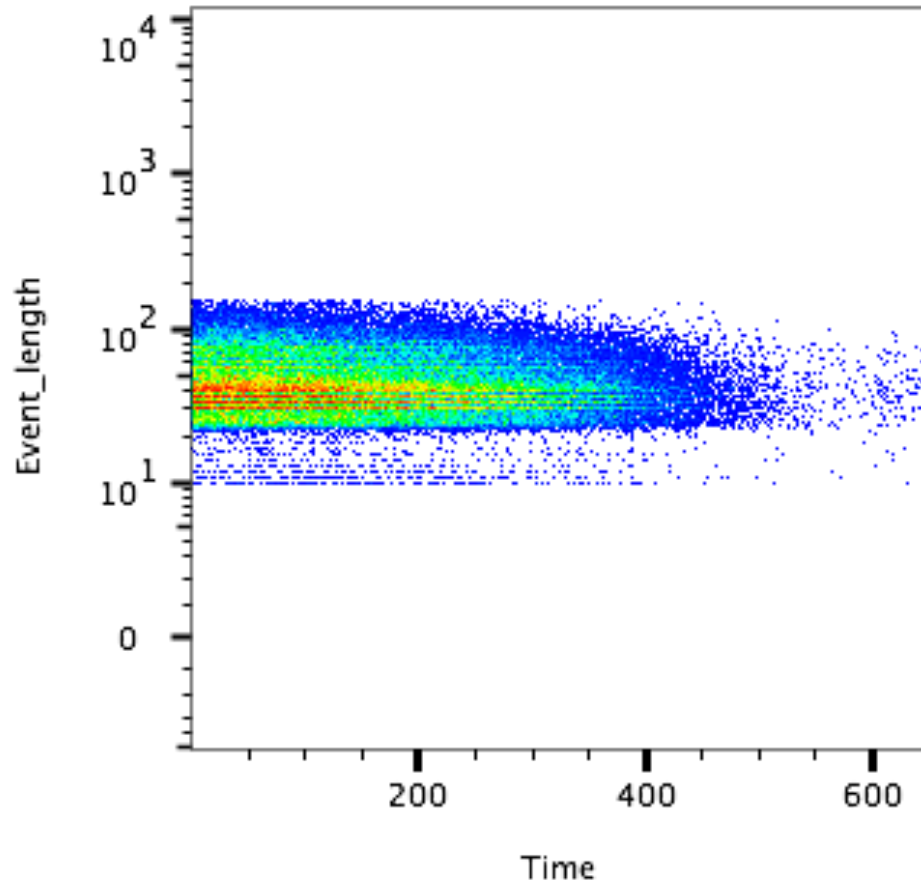


Not so good



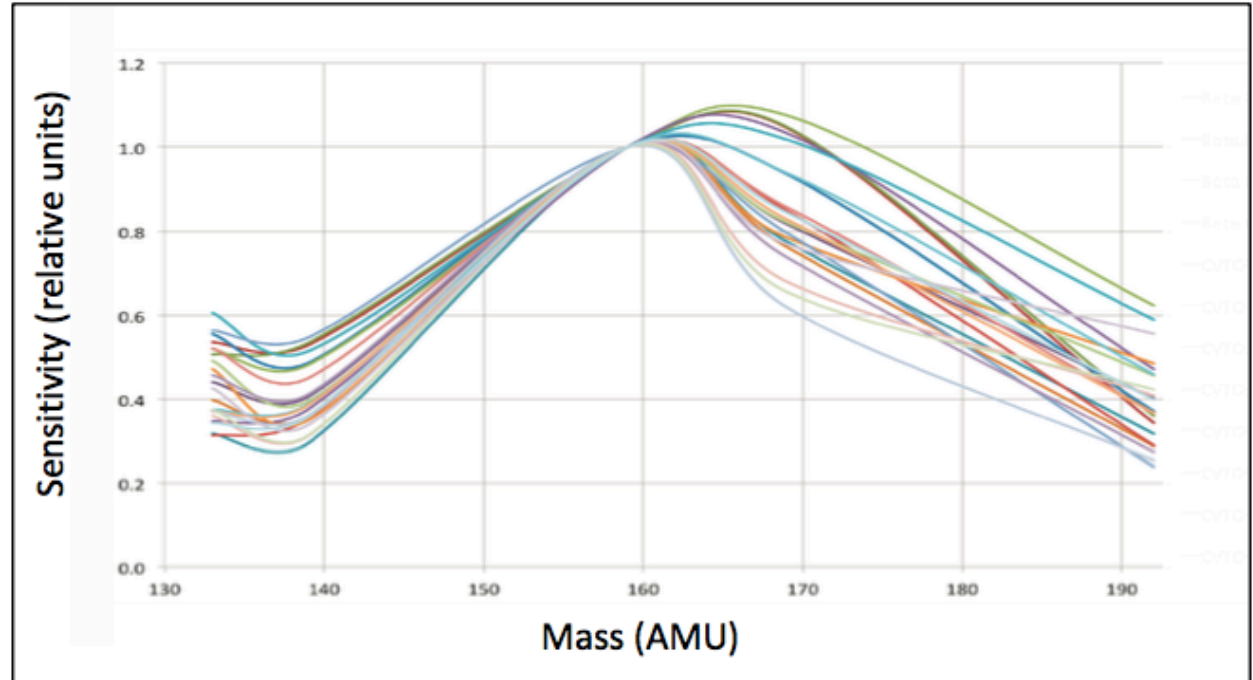
Good

Sample Flow



Metal Sensitivity – Panel Design

- Maxpar Panel Designer – web based tool with guides
- An introduction to mass cytometry: fundamentals and applications - Cancer Immunol Immunother (2013) 62:955–965



- We often recommend starting with a Fluidigm pre-set panel as a base/scaffold
 - Adding additional markers is fairly straightforward

Input: Sample Prep for the CyTOF

Single cell suspensions

- Sample preparation is critical.
 - A little more than “just” flow staining
- As free of debris and clumps as possible.
 - No scatter parameters for gating***
 - Every particle contained within the sample can contribute to signal
 - Filter immediately prior to running
 - Count immediately prior to running
 - Using a method that allows detection of debris
- Beware the water washes
 - Not a bad idea to seek help when attempting this.

Input: Sample Introduction

- Maximum 5 pushes/hr – each push = 10min + washing
- Maximum ~ 500 events per second during run.
- Maximum $\sim 2.5 \times 10^6$ cells run per hour

Begin with 2×10^6 cells: after staining/washing you have $\sim 1 \times 10^6$ remaining: resuspend in 1mL and do two pushes: ~ 30 minutes of run time: recover $\sim 2 \times 10^5$ cells

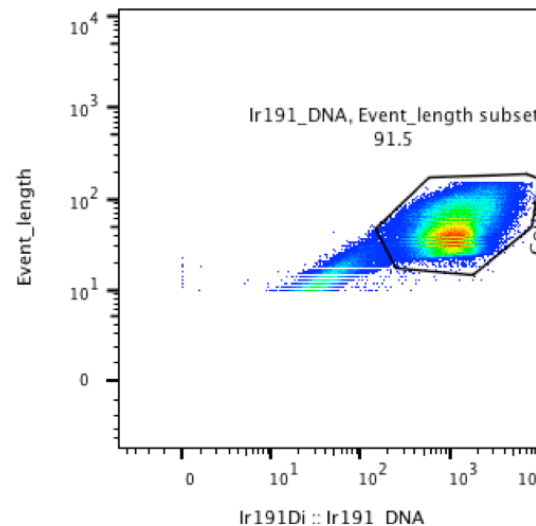
- Can you utilize the barcoding technologies – Palladium or CD45
 - Improves sample throughput for smaller cell numbers
 - Improves standardization of data over time
 - Easy to set up and use

Output: Sample Recovery

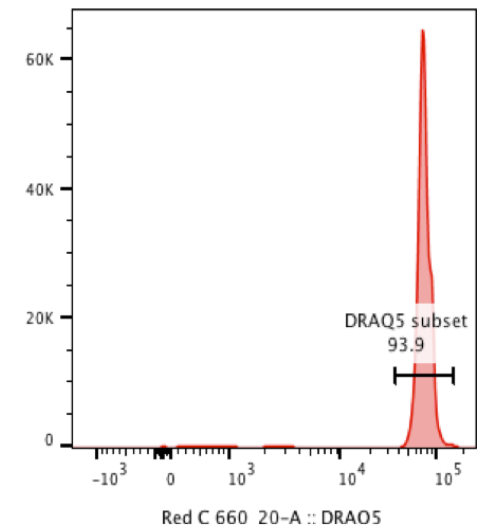
- Suggested as $\sim 30\%$ recovery
 - Increased in the Helios
- Our numbers suggest between 20-30% in our system

- Recovery Comparison
 - CyTOF: $\sim 22\%$
 - LSR: $\sim 31\%$
 - FACSCanto: $\sim 50\%$

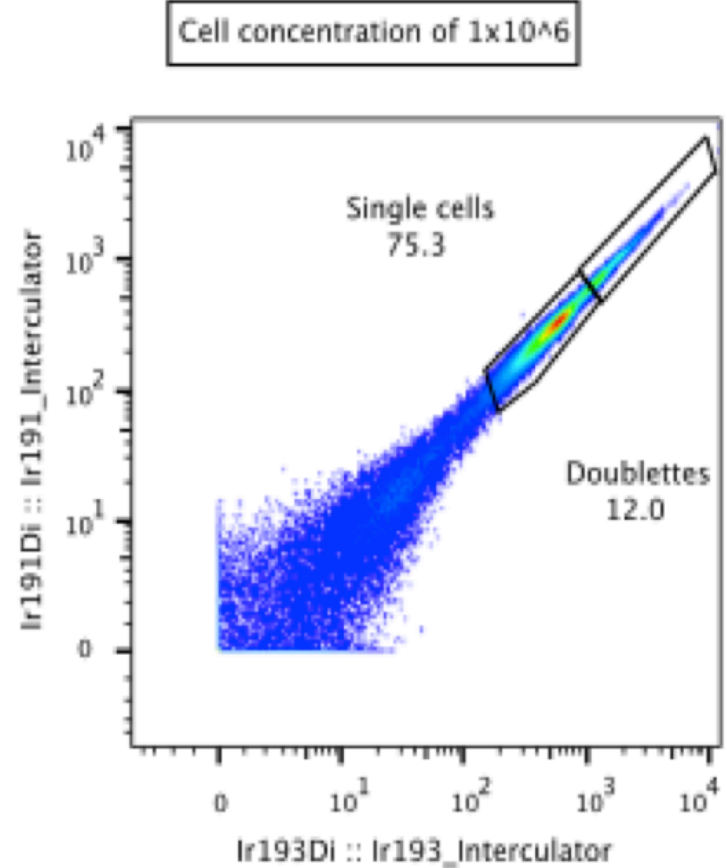
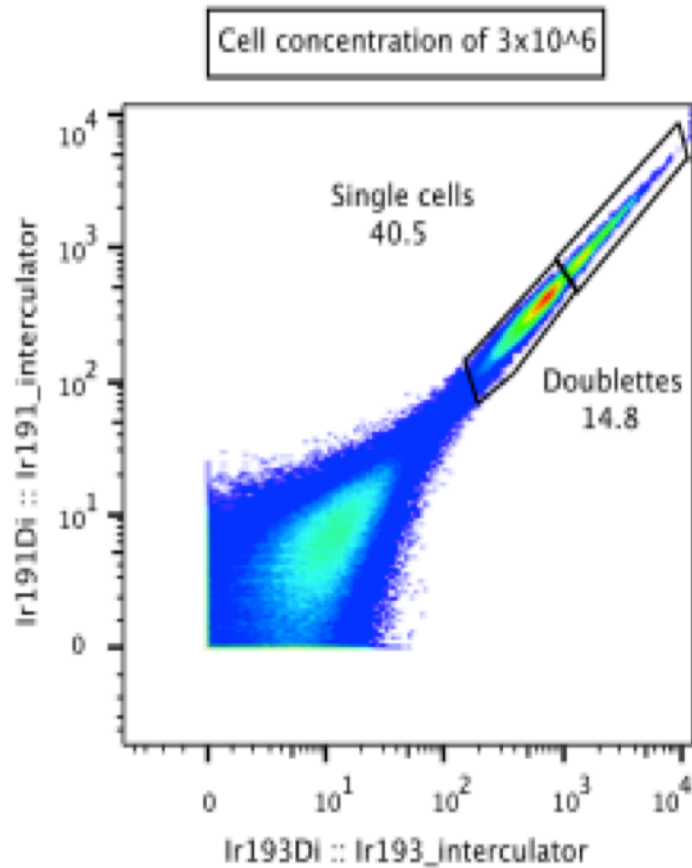
CyTOF data



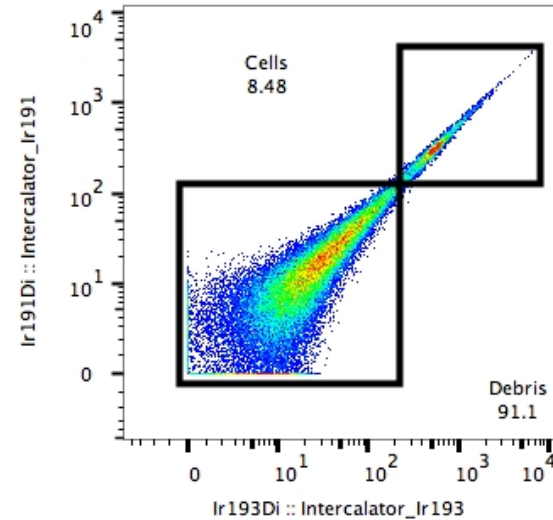
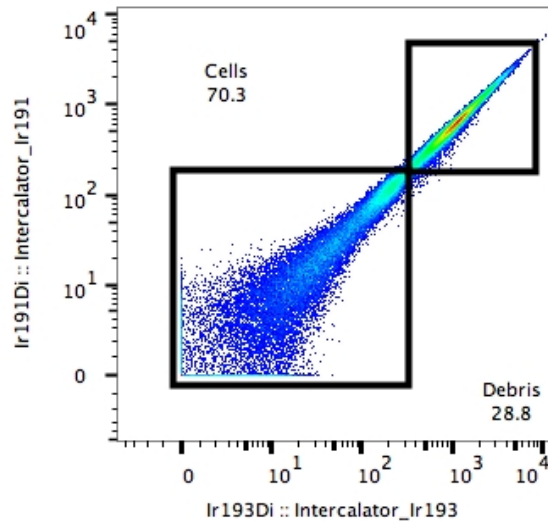
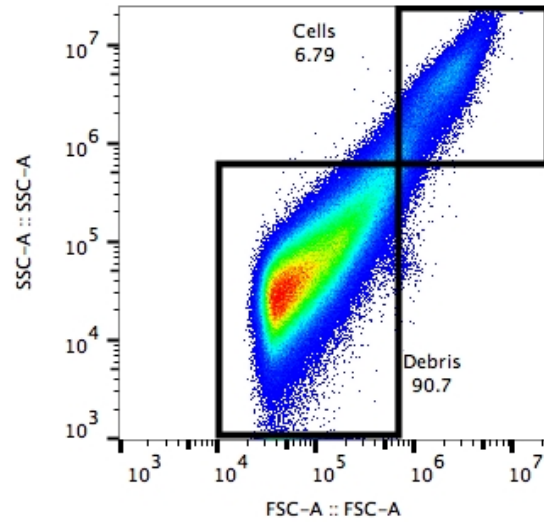
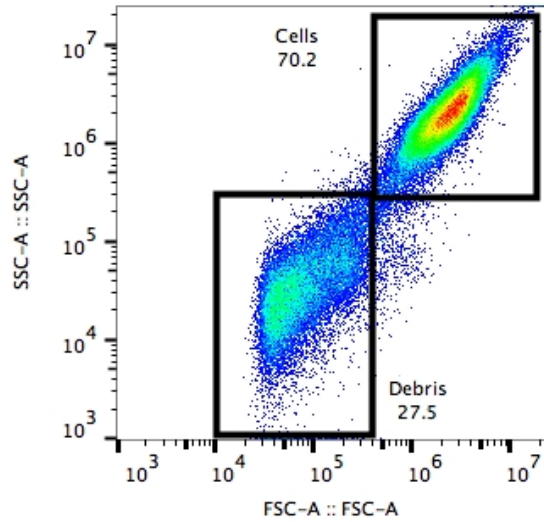
LSRII data



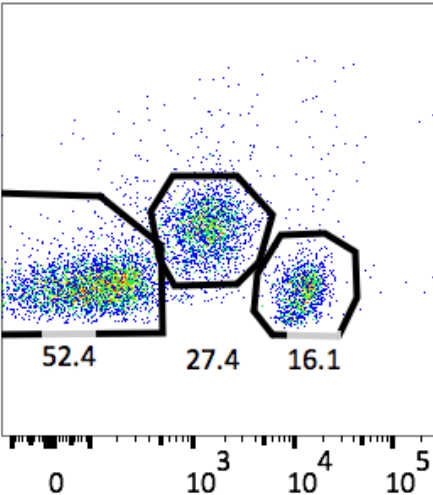
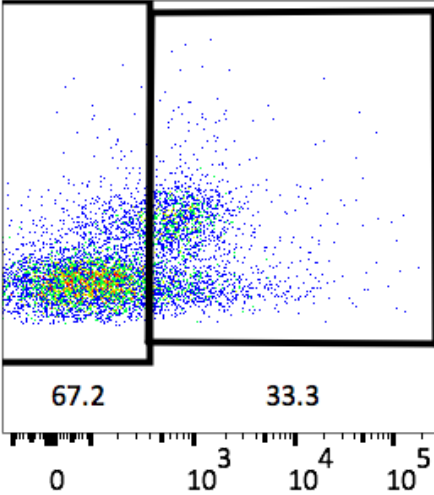
Sample Concentration



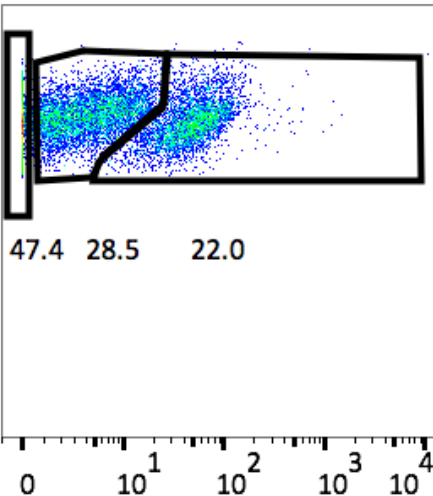
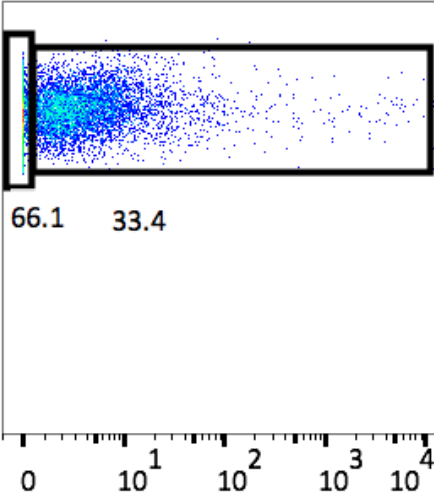
Sample Cleanliness



Output: Comparing the Signal



Flow Run



CyTOF Run

CD279

CD4