

Musculoskeletal Development

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IND464: Musculoskeletal Basic Science

Fall 2019

Musculoskeletal System

- **Bone**

- 206 bones in the adult human skeleton
- Major functions:
 - Mechanical support
 - Protects organs
 - Provides environment for hematopoiesis
 - Stores minerals, growth factors, and hormones

- **Cartilage**

- 3 types: hyaline, elastic, and fibrocartilage
- Provides low friction surface for motion and withstands tensile, shear, and compressive forces

- **Skeletal Muscle**

- Provides contractile forces to move the bones of the skeleton
- Maintains body temperature

- **Tendon**

- Attach muscle to bone and transmit contractile forces from muscle to bone

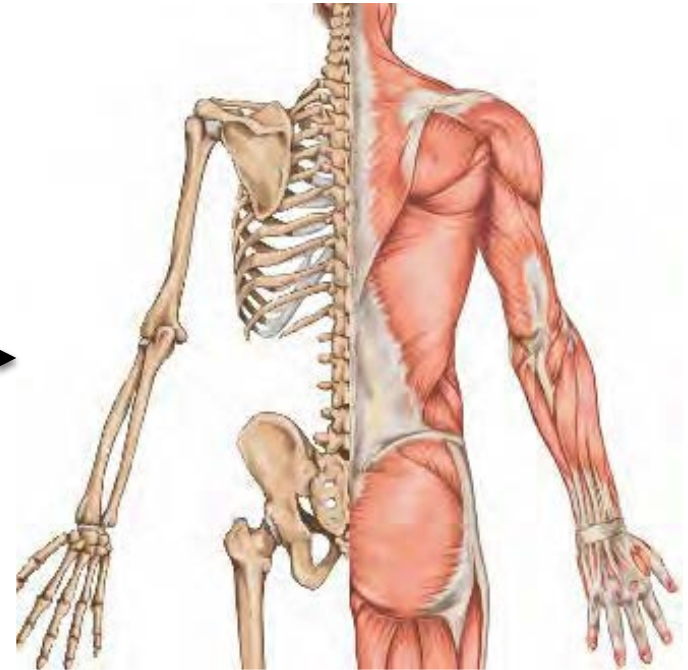
- **Ligament**

- Attach bone to bone



<https://cpd.cqu.edu.au/course/info.php?id=456>

Musculoskeletal System



<https://cpd.cqu.edu.au/course/info.php?id=456>

Lecture Objectives

- Introduction to limb patterning and the signaling centers/pathways responsible for proximal-to-distal and anterior-to-posterior patterning
- Introduction/review of intramembranous and endochondral bone formation and mesenchymal progenitor cell differentiation
- Introduction to joint formation and gene expression patterns characteristic of a developing joint

Embryogenesis/Organogenesis

Mouse E3.5
Human Day 5

E6.5-7.0
Week 2-3

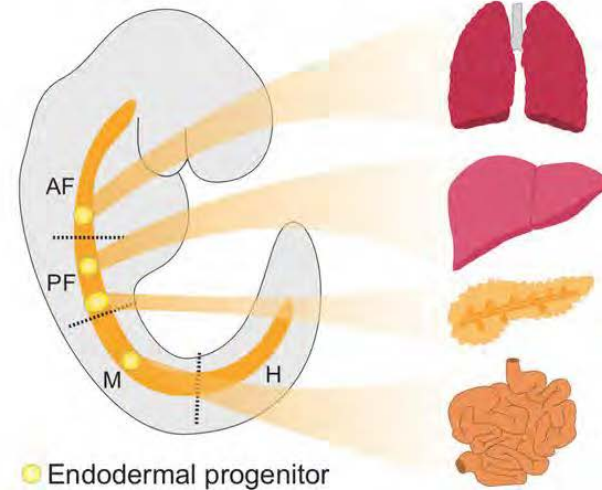
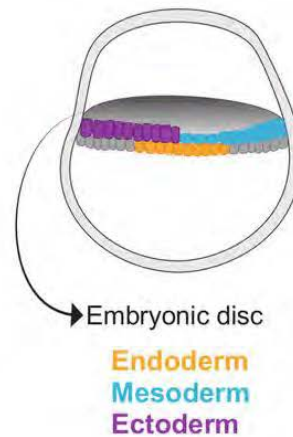
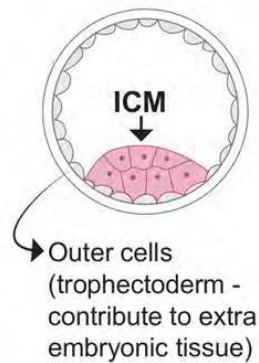
E8.5-9.0
Week 4

Blastocyst

Gastrula

Early organogenesis

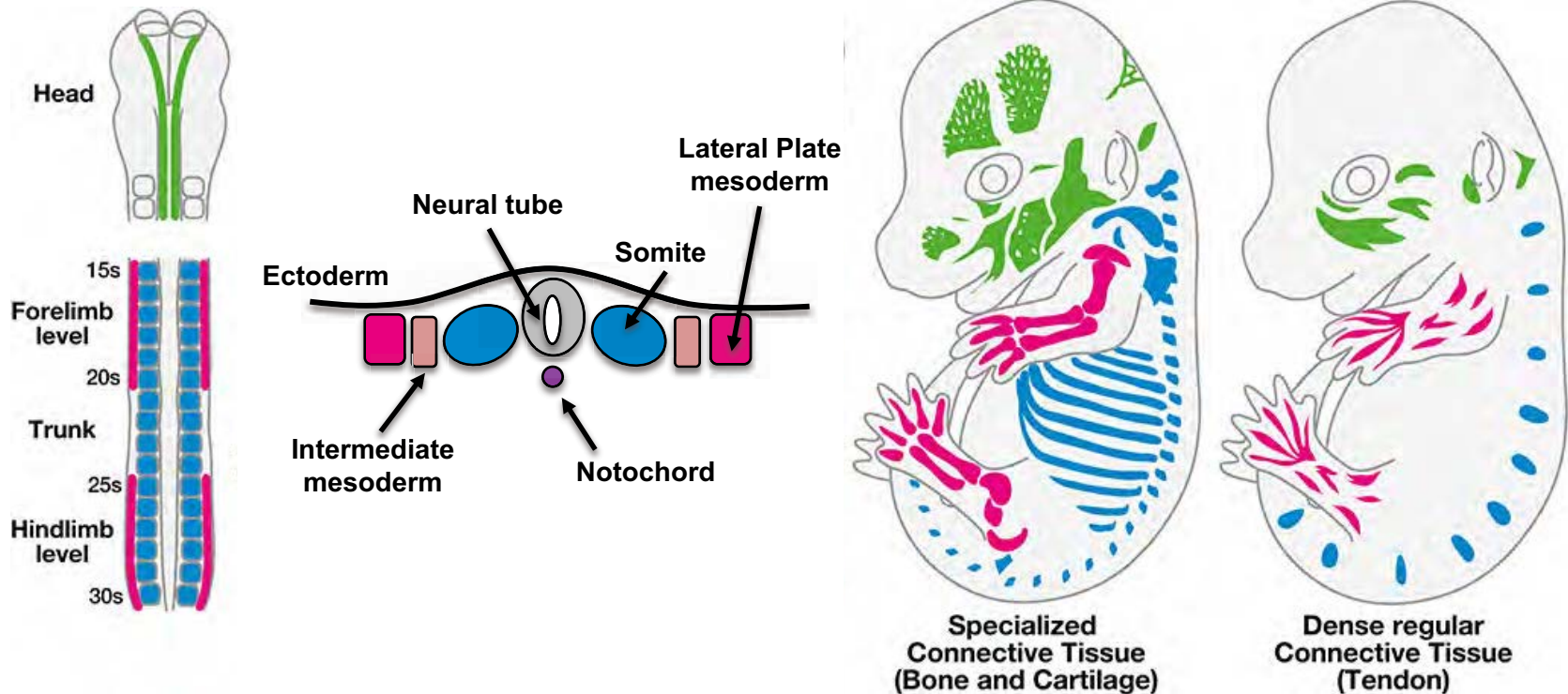
Adult



Endoderm: respiratory system, digestive system
Mesoderm: musculoskeletal system, circulatory system
Ectoderm: skin, nervous system

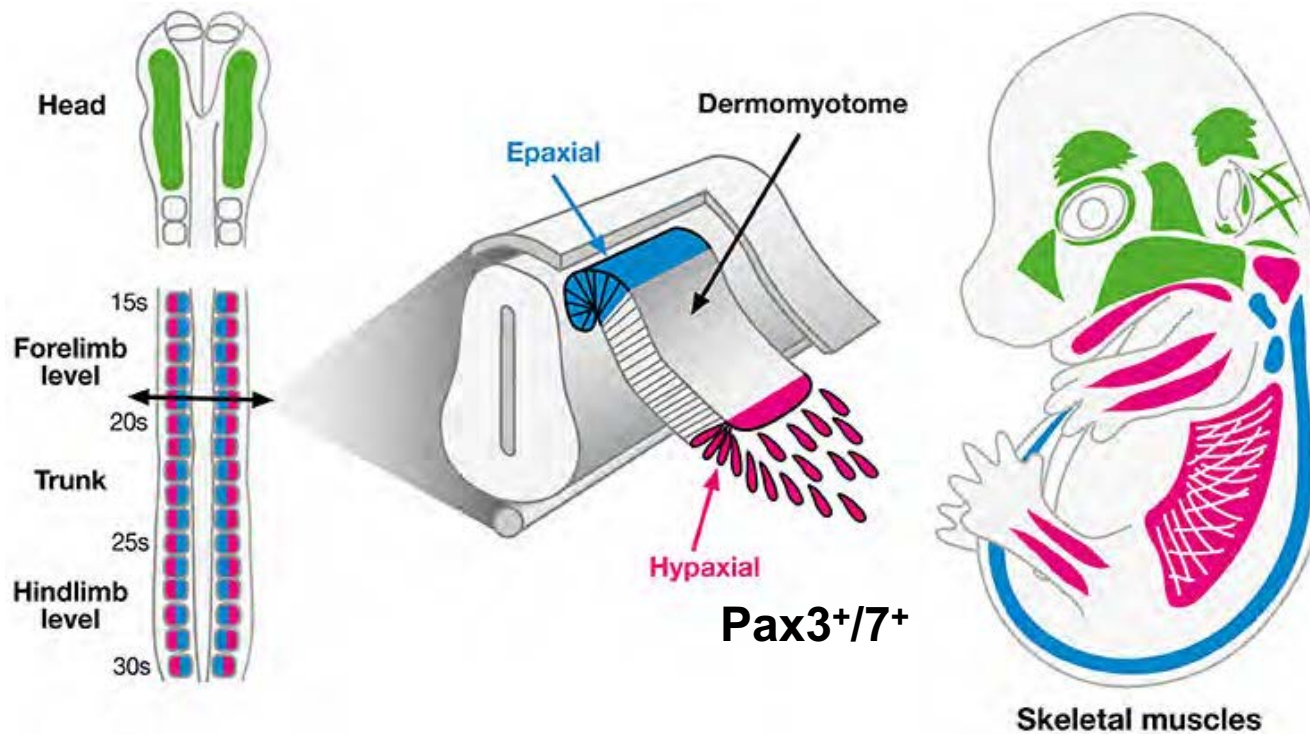
Prior, et. al. (2019) *Gut* doi:10.1136/gutjnl-2019-319256.

Embryonic Origins of Musculoskeletal Tissues



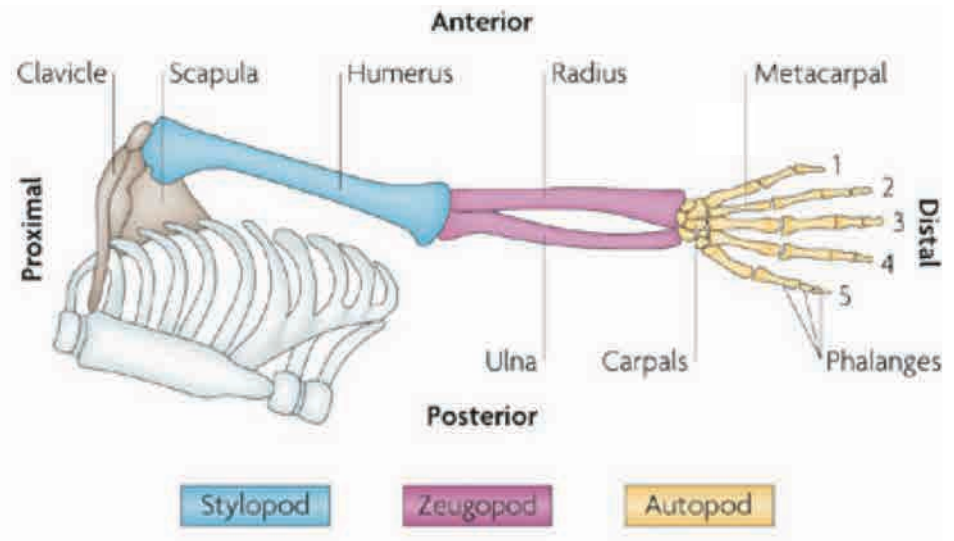
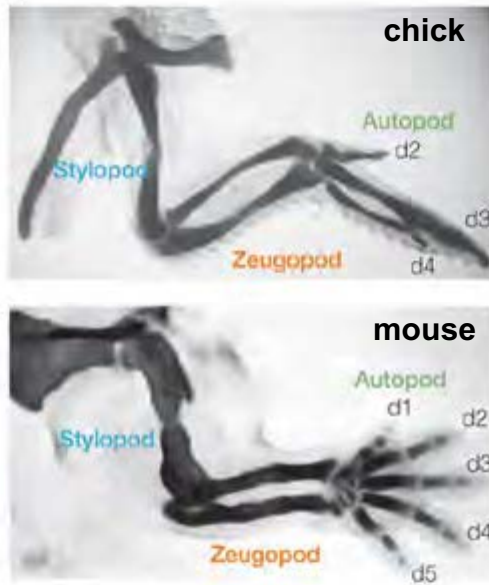
Adapted from Nassari et. al. (2017) *Front Cell Dev Biol.* 5:22.doi: 10.3389/fcell.2017.00022.

Embryonic Origins of Musculoskeletal Tissues



Adapted from Nassari *et. al.* (2017) *Front Cell Dev Biol.* 5:22.doi: 10.3389/fcell.2017.00022.

Patterning of the Limb Skeleton



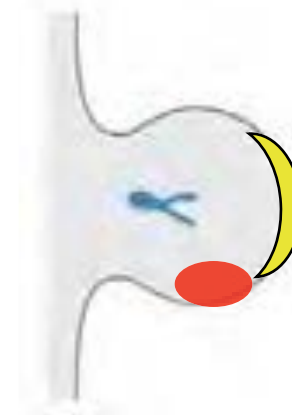
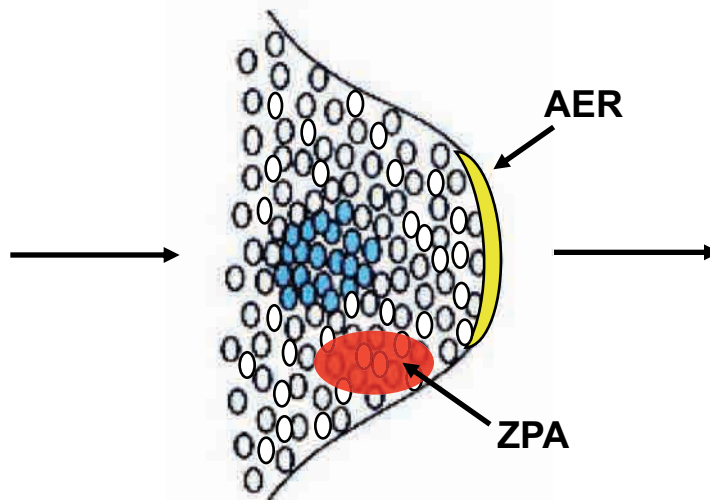
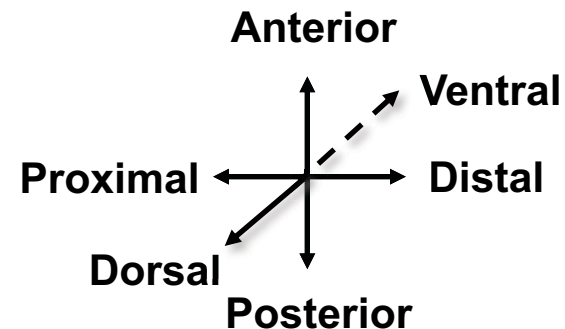
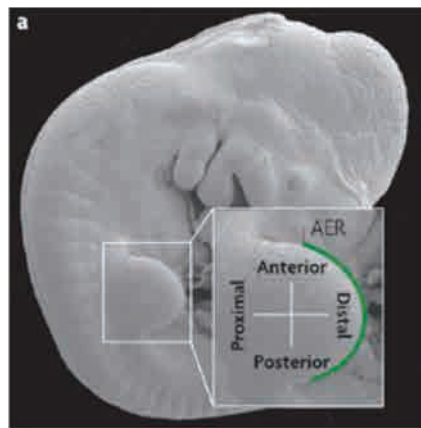
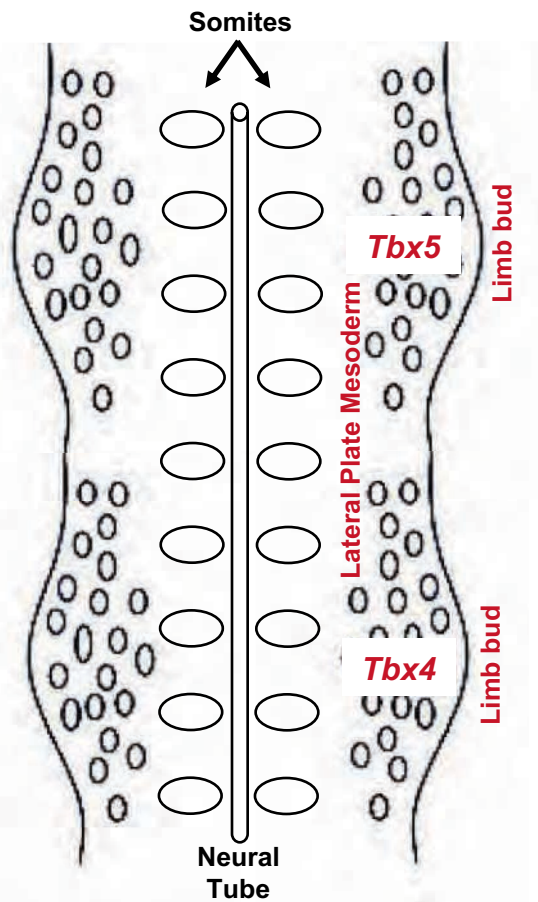
Patterning: the process in which the positions and identities of cells with different fates are laid down.

Specification: the process preceding determination during which a cell acquires its fate. The exposure of specified cells to different signals might alter their fates; the fate of specified cells is not fixed.

Determination: When cell fate is fixed so that the cell will initiate differentiation into the specified cell type even if the cell is isolated or transplanted into a different environment or tissue; occurs before the appearance of cell-type-specific morphological characteristics, but is often followed by differentiation.

Zeller, *et. al.* (2009) *Nat Rev Gen.* 10: 845-858.
Niswander (2003) *Nat Rev Gen.* 4: 131-142.

Limb bud Development, Patterning, and Chondrogenesis



Mouse: E9.5-E10.0

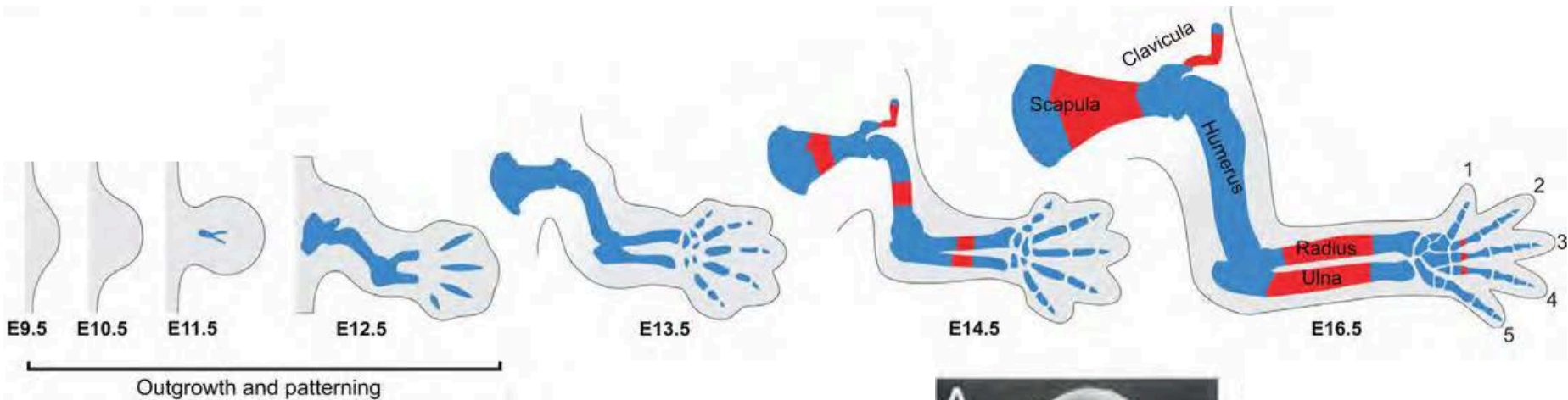
Human: 4-5 weeks

E10.5-E11.5

E11.5-E12.5

Adapted from Ornitz and Marie (2002) *Genes and Development*. 16: 1446-65.
Zeller, et. al. (2009) *Nat Rev Gen.* 10: 845-858.

Mouse Limb Patterning and Development

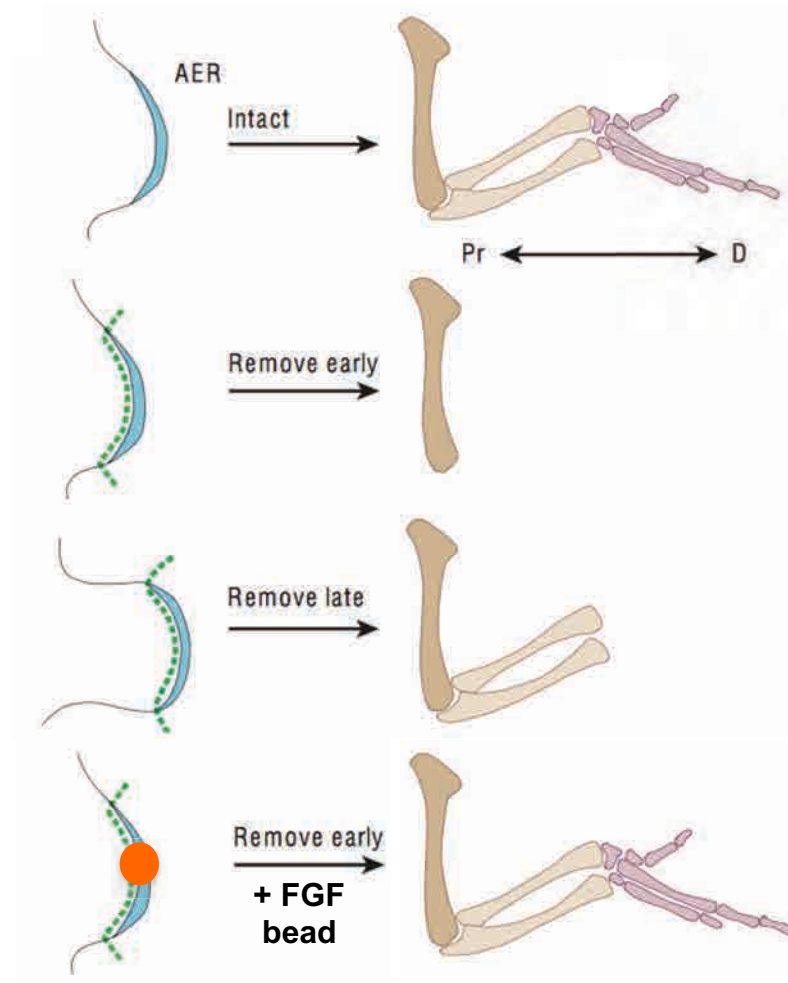
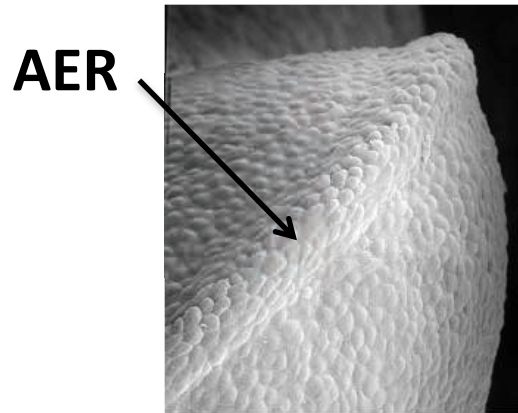


- *Hindlimb development is delayed by 0.5 days*



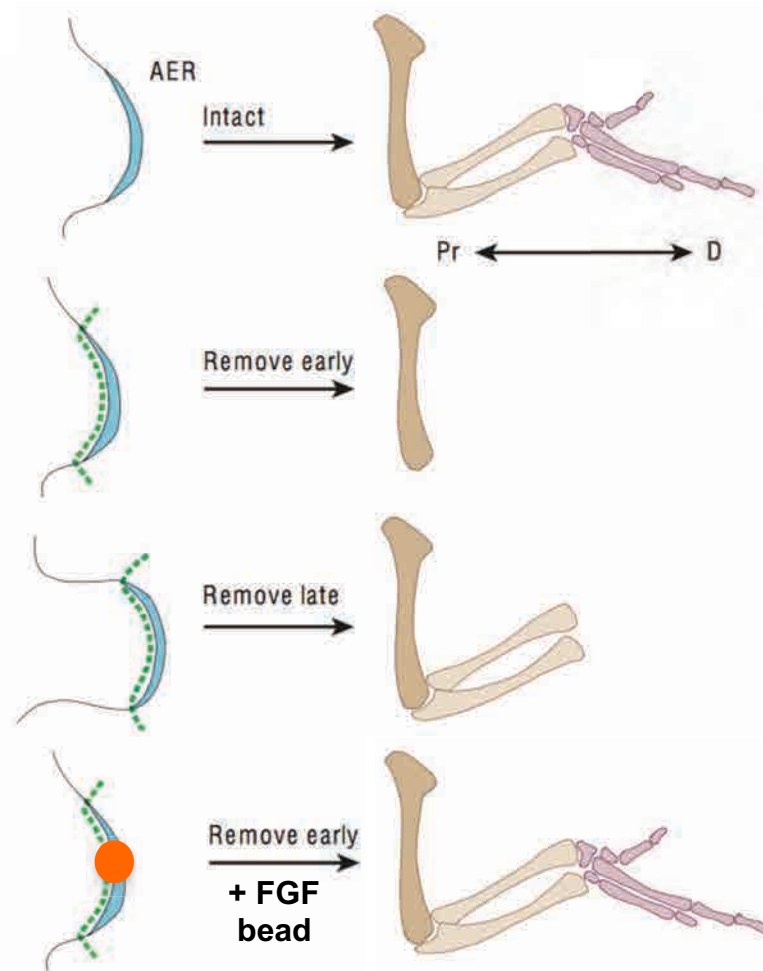
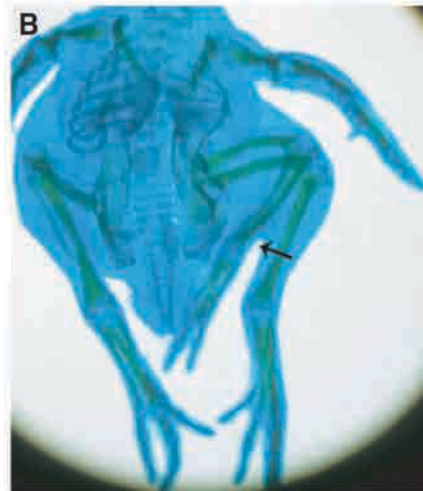
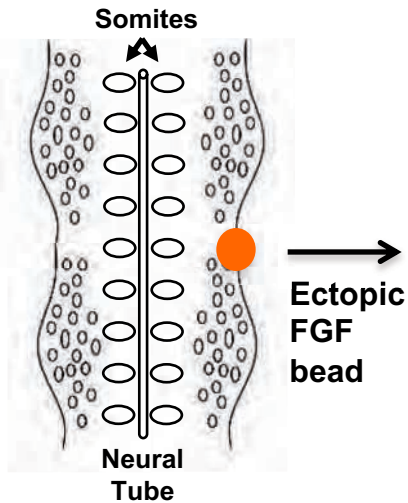
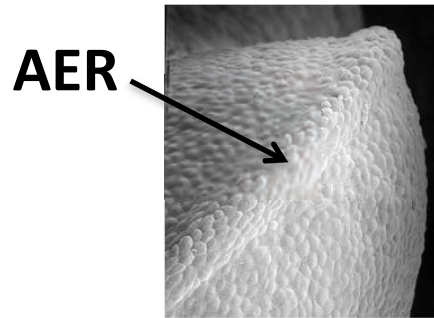
Adapted from Zuniga (2015) *Development*. 142: 3810-20.

Influence of the Apical Ectodermal Ridge (AER) on Proximodistal Limb Patterning



Adapted from Mariani and Martin (2003) *Nature* 423: 319-325.

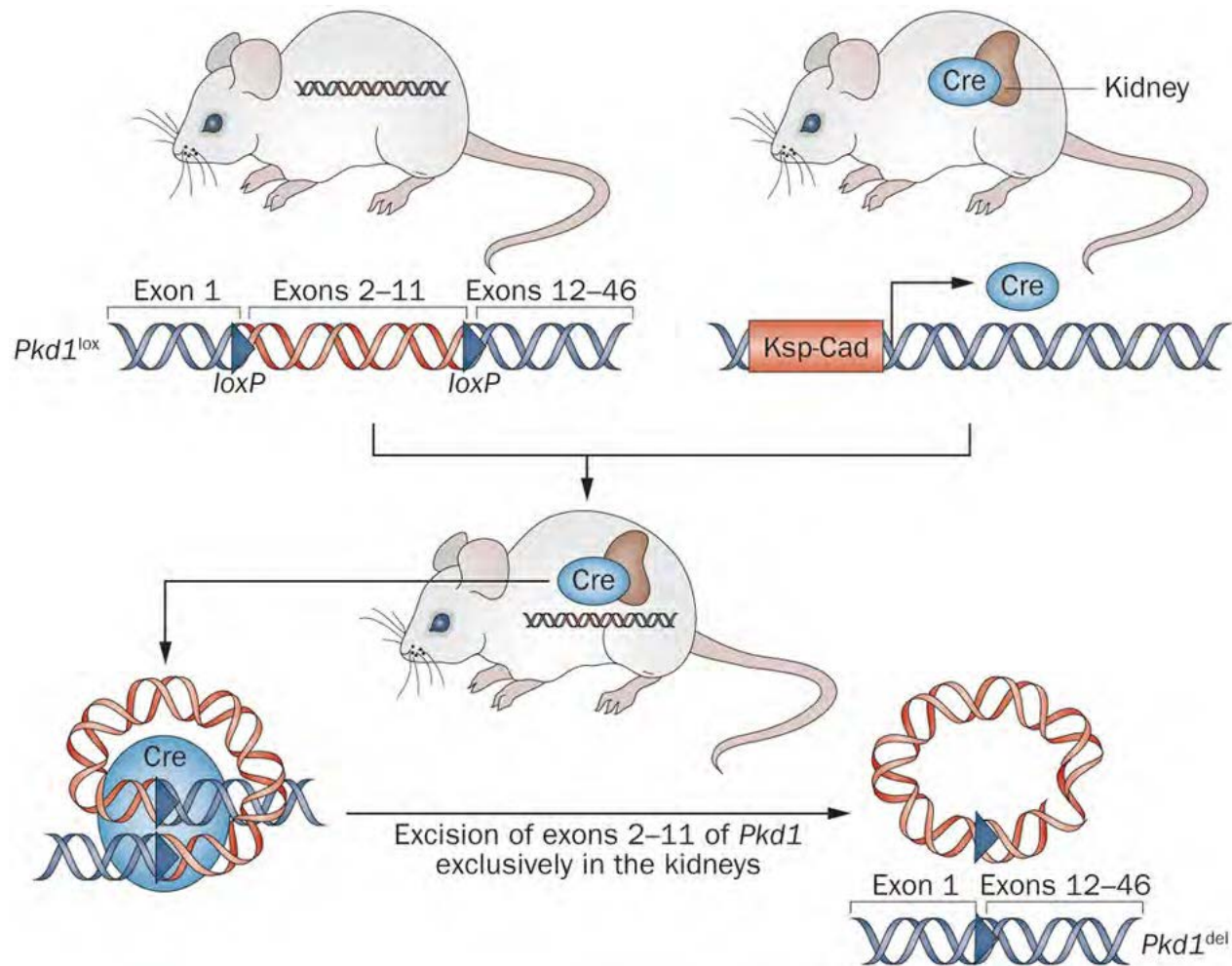
Influence of the Apical Ectodermal Ridge (AER) on Proximodistal Limb Patterning



Adapted from Tickle (2015) *J. Anat.* 227: 418-430.

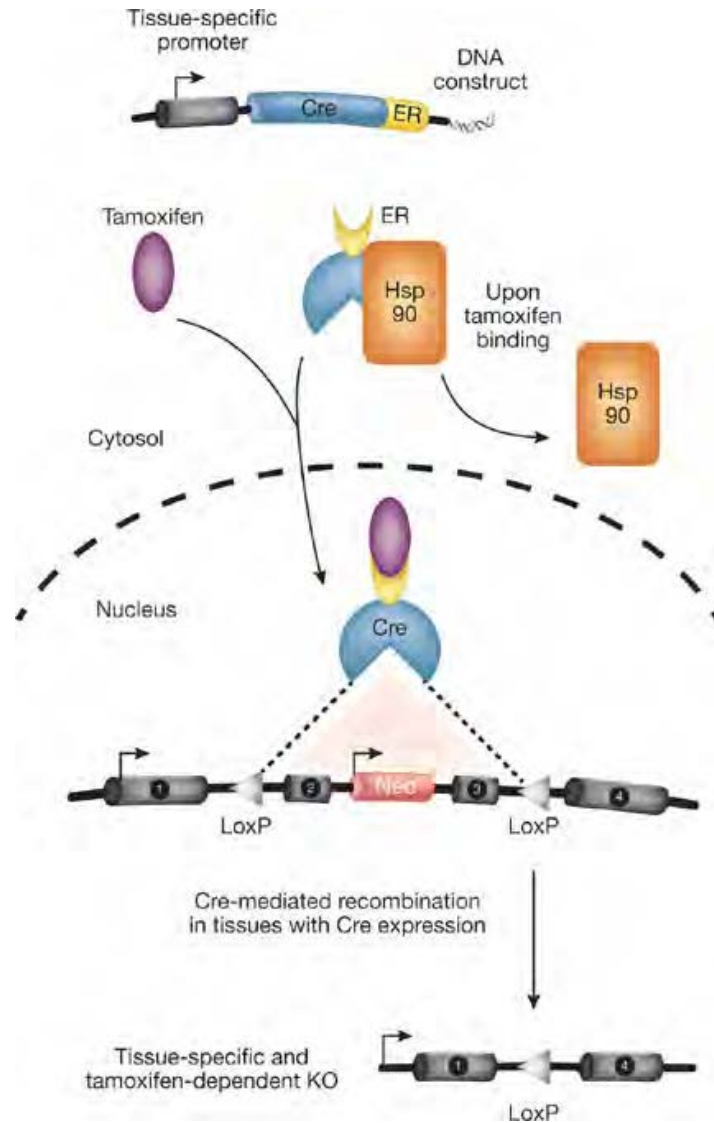
Adapted from Mariani and Martin (2003) *Nature* 423: 319-325.

Overview of the Cre-loxP Recombination System



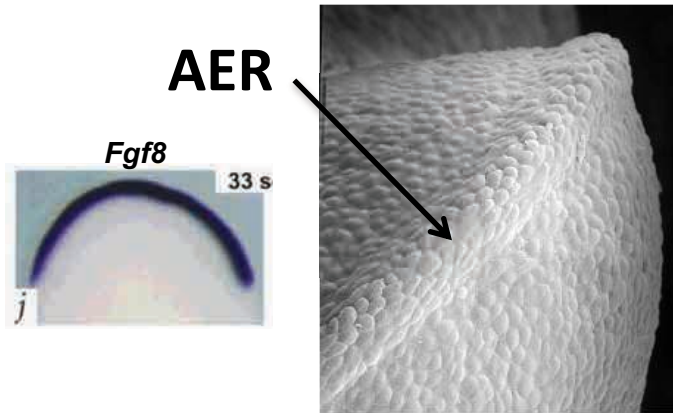
Happé, H. and Peters, D. J. M. (2014) *Nat. Rev. Nephrol.* doi:10.1038/nrneph.2014.137

The Tamoxifen-inducible Cre-loxP System



Gunschmann, C. (2014) *J Invest Dermatol.*
doi:10.1038/jid.2014.213

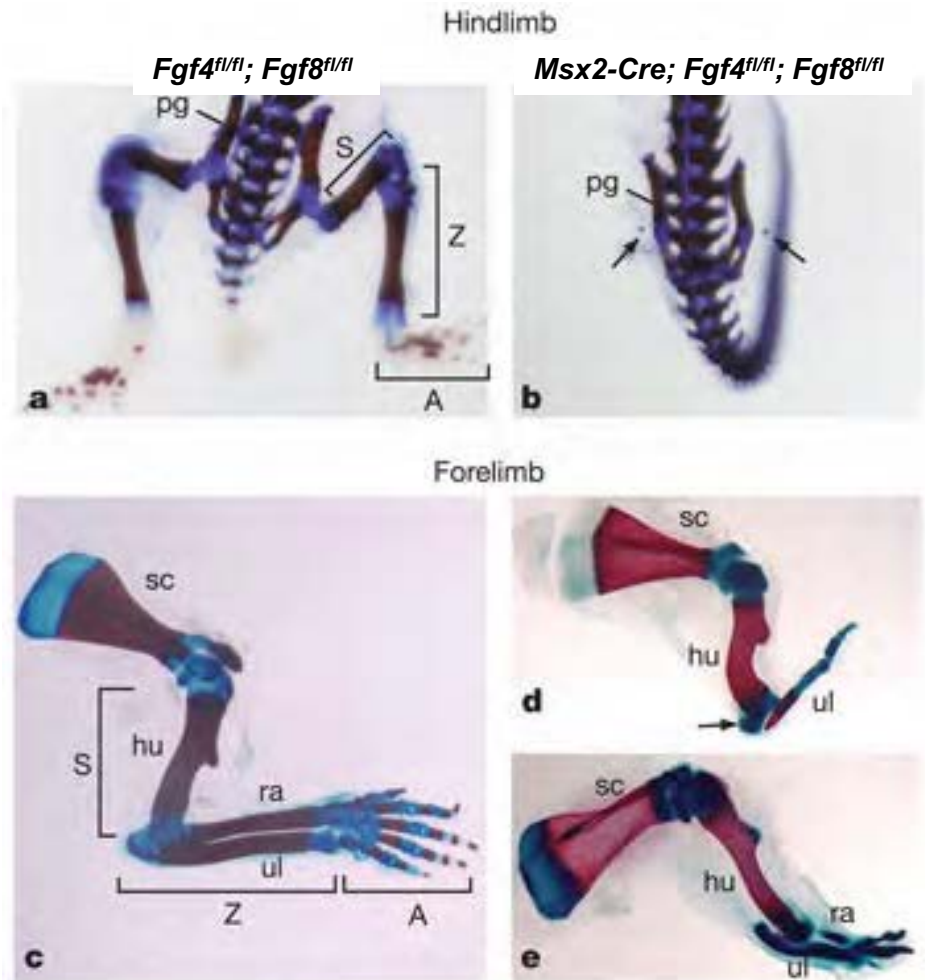
Influence of the Apical Ectodermal Ridge (AER) on Proximodistal Limb Patterning



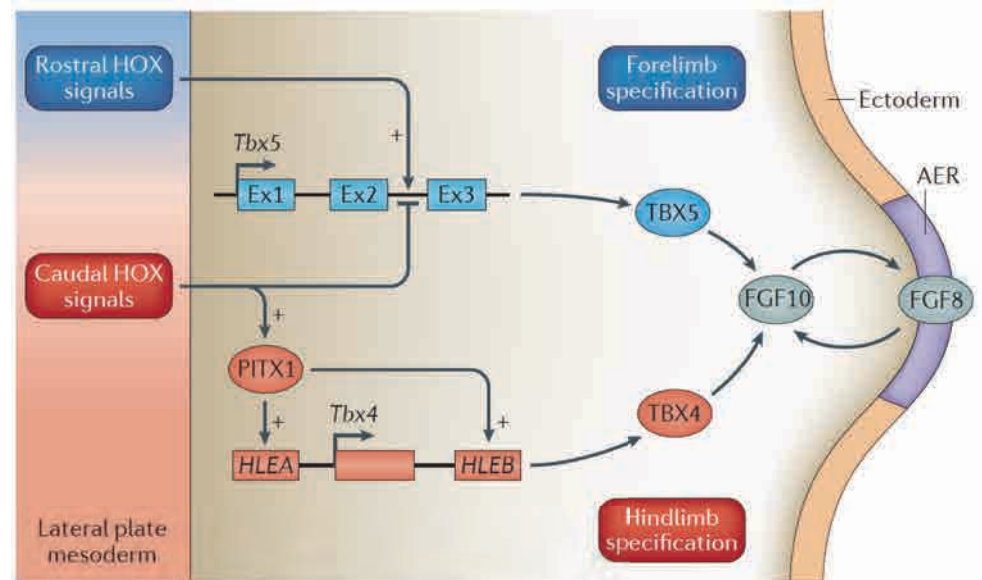
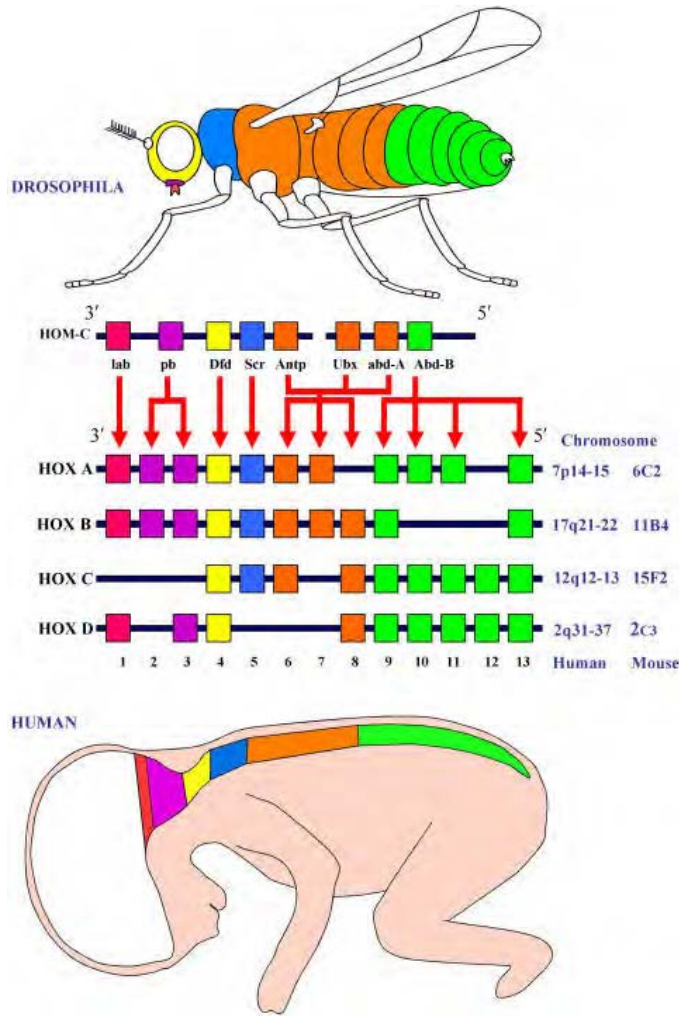
- FGF4, 8, 9, and 17 are all expressed in the AER.
- Only *Fgf8* is essential for normal limb bud development.
- Deletion of both *Fgf4* and *Fgf8* from the AER completely disrupts limb bud development.

Lewandoski, et. al. (2000) *Nat Gen* 26: 460-463.

Sun, et. al. (2002) *Nature* 418: 501-508.

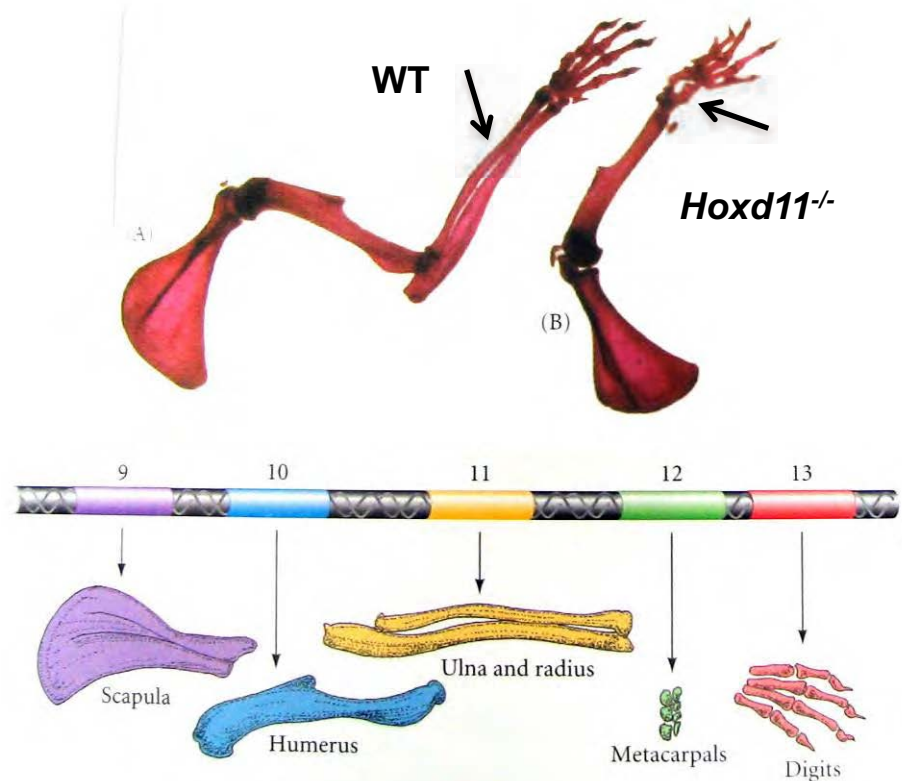
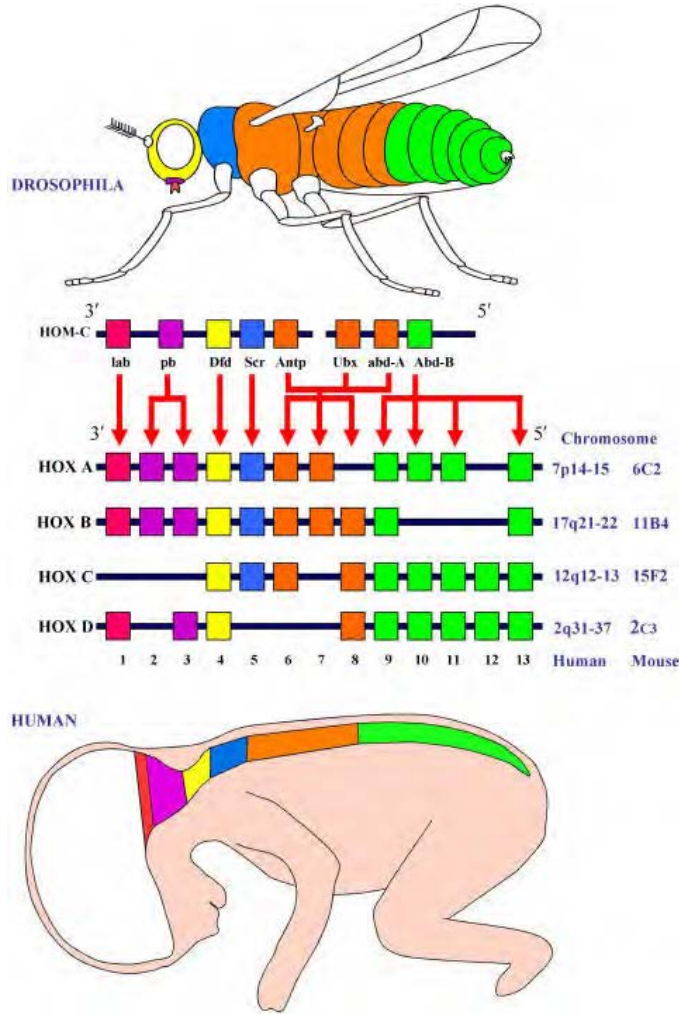


Hox Gene Regulation of Proximodistal Patterning



Lappin, et. al. (2006) *Ulster Med* 75: 23-31.
 Petit, et. al. (2017) *Nat Rev Gen*, ePub.

Hox Gene Regulation of Proximodistal Patterning



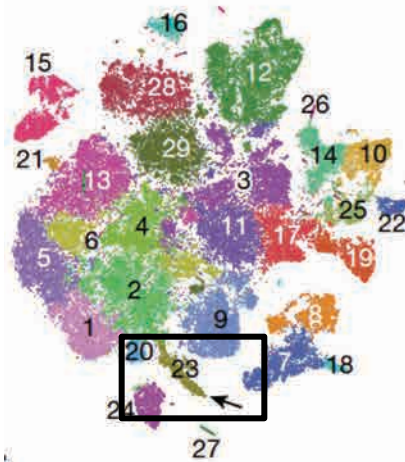
Lappin, et. al. (2006) *Ulster Med* 75: 23-31.
 Davis, et. al. (1995) *Nature* 375: 791-795.

Identification of Novel AER-associated Genes using scRNA-seq

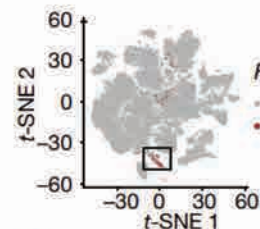
“The single-cell transcriptional landscape of mammalian organogenesis”

a

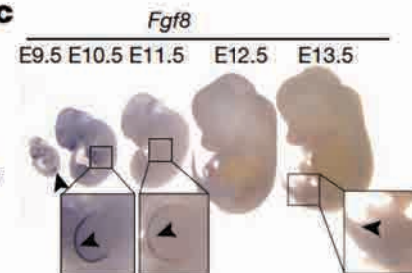
- 1–Keratinocyte (*Col19a1*⁺)
- 2–Epidermal stem cells (*Meis1*⁺)
- 3–Branchial arch ectodermal cells
- 4–Epidermal progenitors (*Igf1bp2*⁺)
- 5–Keratinocyte (*Brinp1*⁺)
- 6–Keratinocyte (*Krt1*⁺)
- 7–Otic vesicle epithelium
- 8–Otic sensory epithelium
- 9–Pericardium
- 10–Intestinal epithelium
- 11–Second branchial arch epithelium
- 12–Olfactory epithelium
- 13–Hair follicle stem cell
- 14–Intestinal stem cells
- 15–Renal epithelium
- 16–Retina epithelium
- 17–First branchial arch epithelium
- 18–Utricle and saccule epithelium
- 19–Lung epithelium
- 20–Surface ectoderm
- 21–Epidermal stem cells (*Lgr6*⁺)
- 22–Endocrine cells
- 23–Apical ectodermal ridge (AER)
- 24–Urothelium
- 25–Stomach epithelium
- 26–Intestine epithelium (*Car3*⁺)
- 27–Primordial germ cells
- 28–Doublets
- 29–Doublets



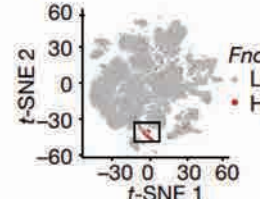
b



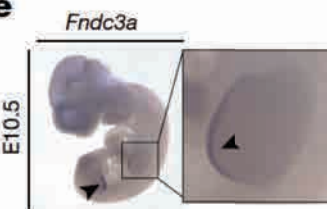
c



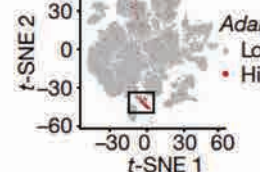
d



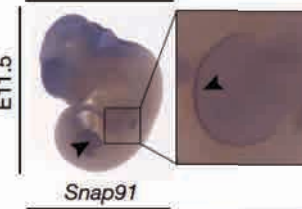
e



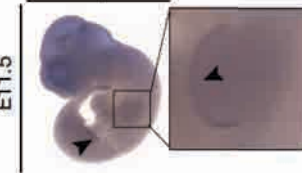
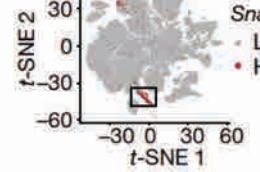
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g

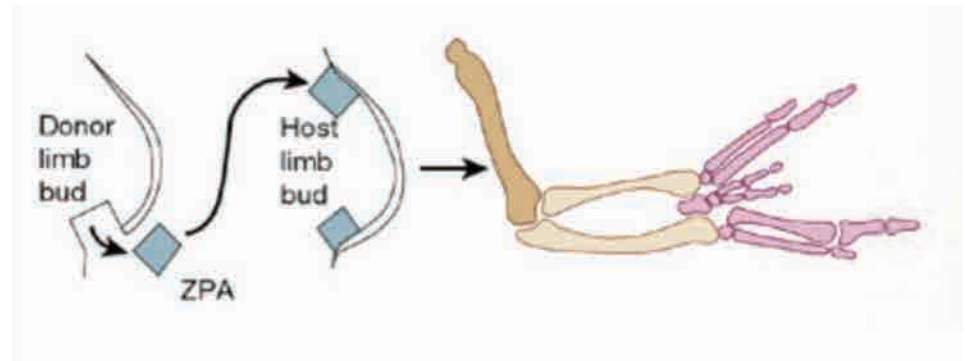
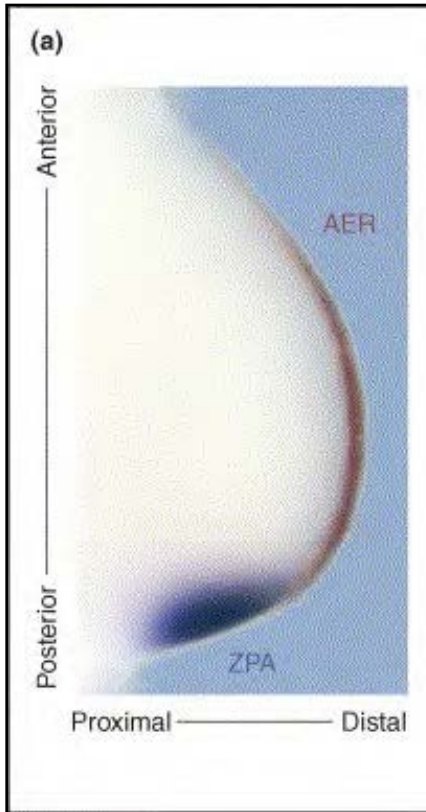


h



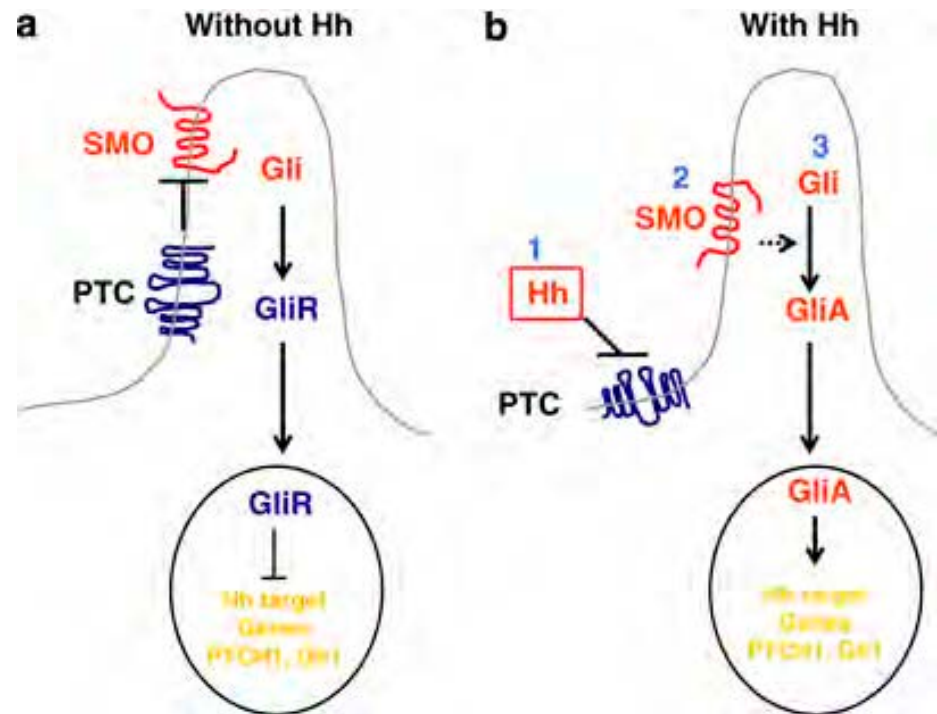
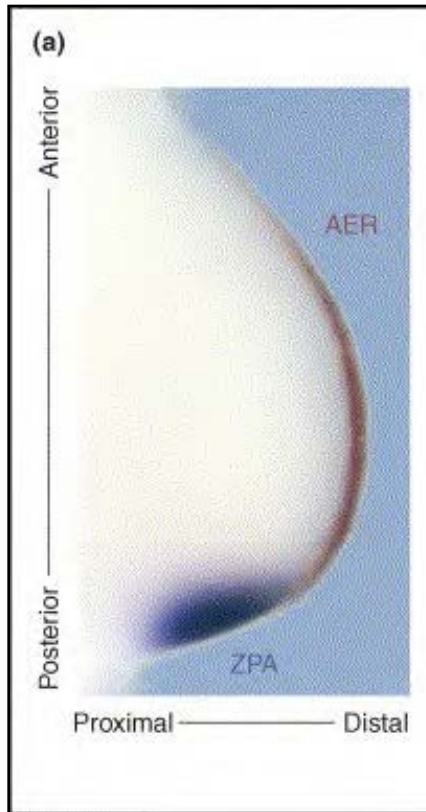
Cao, et. al. (2019) Nature 566: 496-502.

ZPA Regulation of Anterior to Posterior Patterning



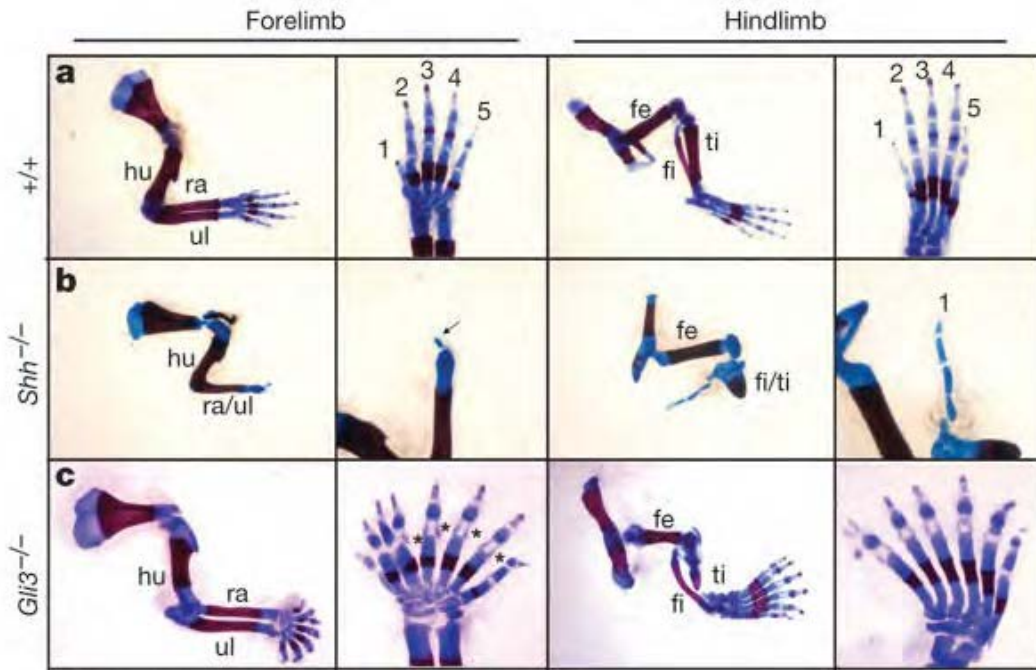
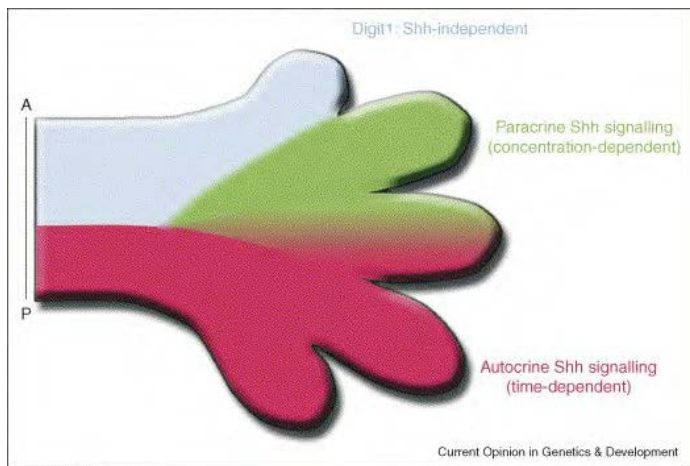
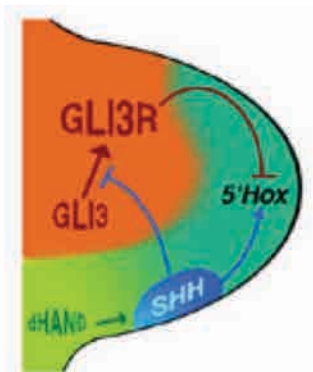
Adapted from Mariani and Martin (2003) *Nature* 423: 319-325.

ZPA/SHH Regulation of Anterior to Posterior Patterning



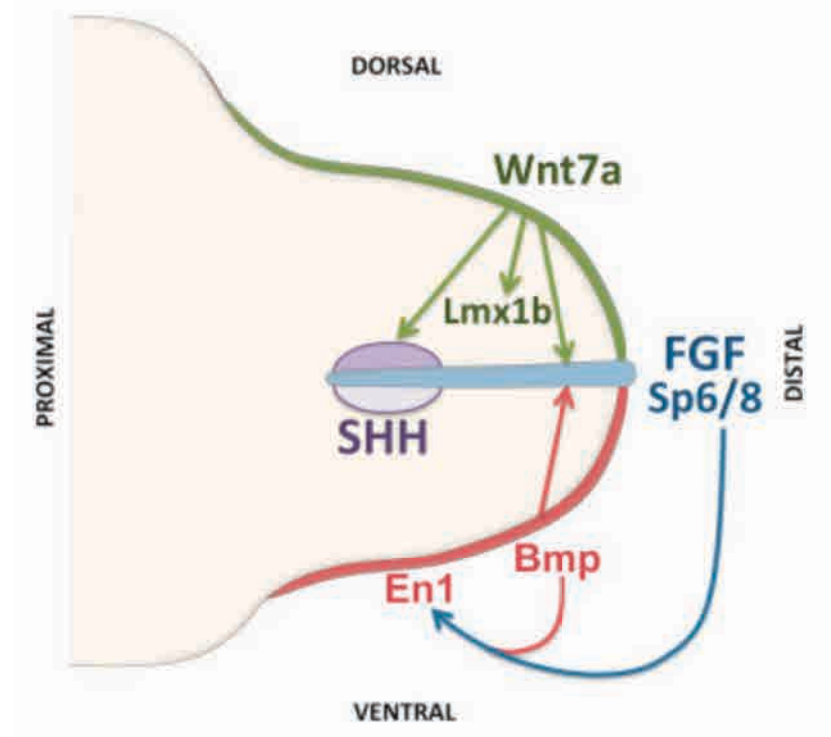
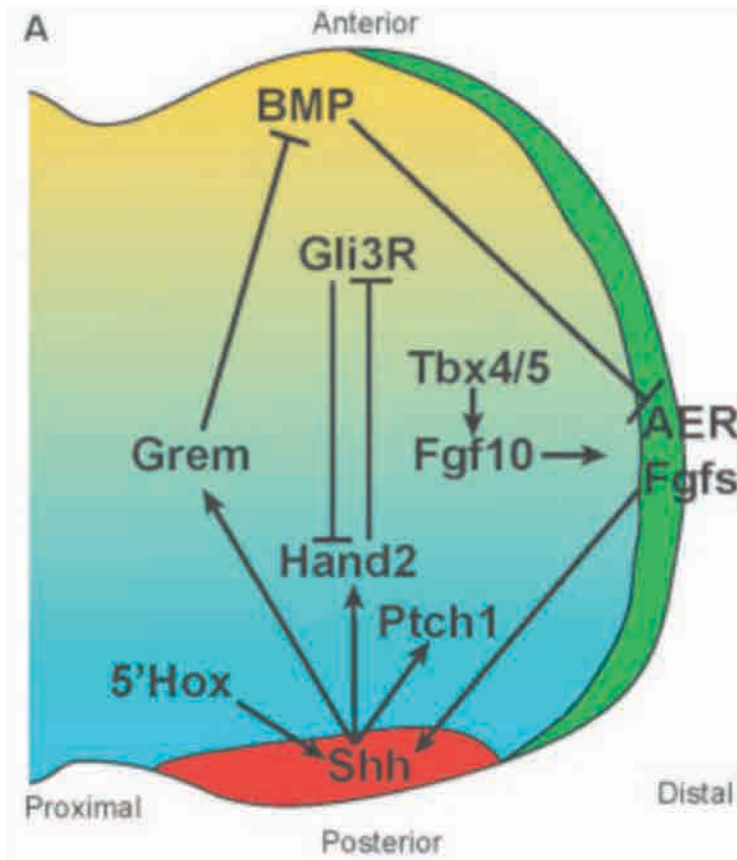
Yang, et. al. (2010) *Oncogene* 29: 469-481.

ZPA (Shh) Regulation of Anterior to Posterior Patterning



Litingtung *et. al.* (2002) *Nature* 418: 979-983.

Coordinated Development of the Vertebrate Limb Across Axes



Young & Tabin (2017) *Dev Biol.* 429: 401-408.
 Delgado & Torres (2017) *Dev Biol.* 429: 382-386.

Human Disorders of Skeletal Patterning, Differentiation, and Growth

- **Polydactyly** (extra digits)
- **Oligodactyly** (missing digits)
- **Amelia** (absence of limbs)
- **Meroamelia** (partial absence of a limb)

- **Brachydactyly** (shortened digits)
- **Campomelic Dysplasia** (*SOX9* haploinsufficiency)
- **Ellis-van Creveld Syndrome** (*EVC* mutations)
- **Cleidocranial Dysplasia** (*RUNX2* haploinsufficiency)

- **Dwarfing Chondrodysplasias**
 - Hypochondroplasia
 - Achondroplasia
 - Thanatophoric Dysplasia



<https://runkle-science.wikispaces.com/Polydactyly>

For review, Kornak and Mundlos (2003) *Am. J. Hum. Genet.* 73: 447-474.

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<http://drpanossian.com/surgical-solutions/hand-deformities/#ectrodactyly>

For review, Kornak and Mundlos (2003) *Am. J. Hum. Genet.* 73: 447-474.

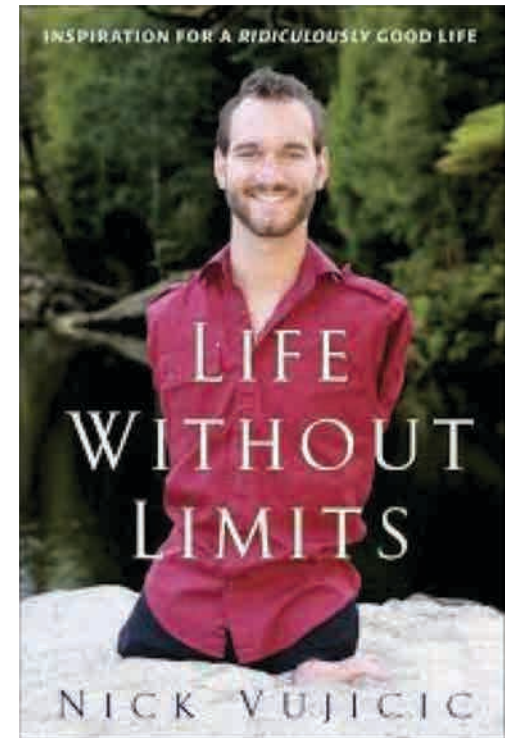
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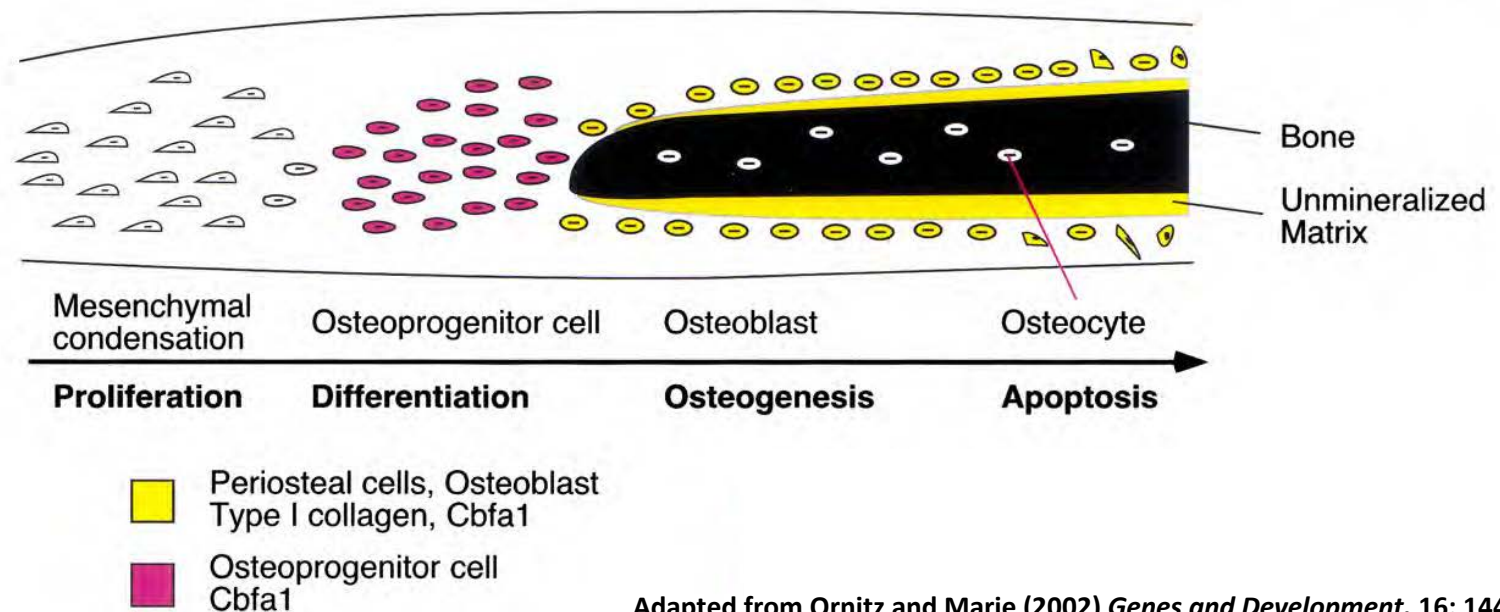
Tetra-amelia (mutations in *WNT3*)



For review, Kornak and Mundlos (2003) *Am. J. Hum. Genet.* 73: 447-474.

Intramembranous *versus* Endochondral Ossification

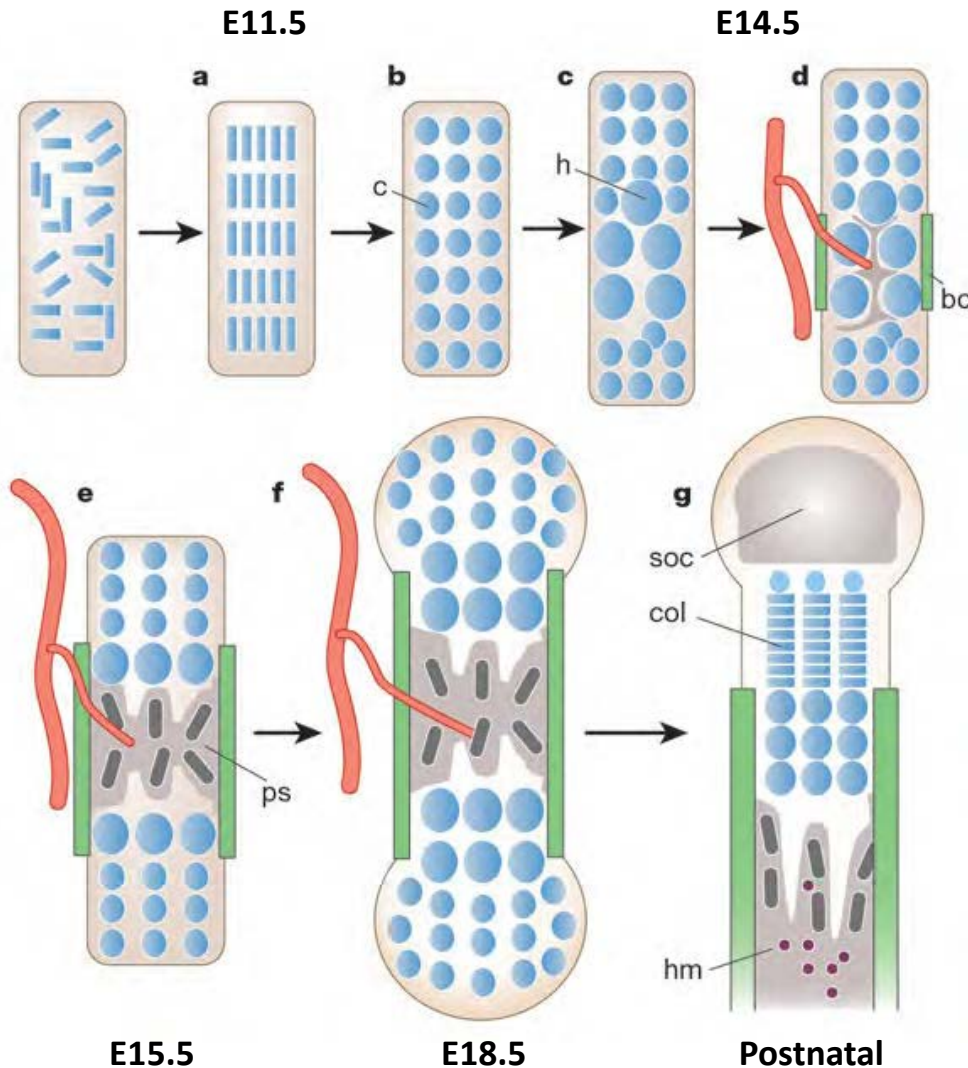
- **Intramembranous:** bone formation directly from mesenchymal condensations; flat bones (skull, mandible, clavicles)



Adapted from Ornitz and Marie (2002) *Genes and Development*. 16: 1446-65.

- **Endochondral:** bone formation following the formation of a hyaline cartilage template; all other bones (long bones, etc.)

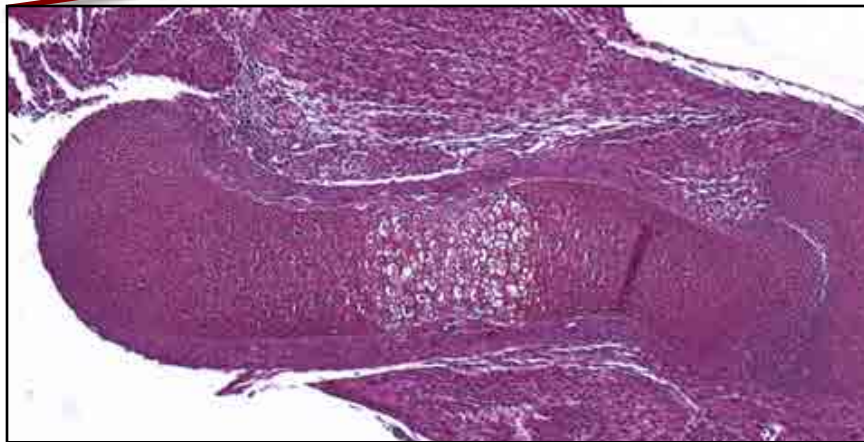
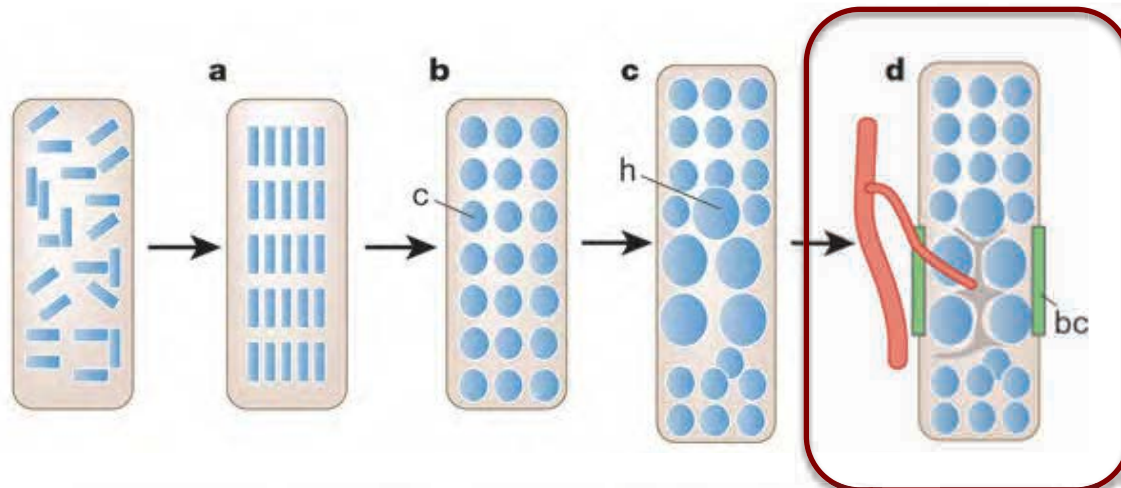
Endochondral Bone Formation



- a. Mesenchymal condensations
- b. Chondrogenesis and chondrocyte proliferation
- c. Onset of hypertrophy
- d. Terminal chondrocyte differentiation
- e. Primary ossification center formation
- f-g. Establishment of epiphyseal growth plates and secondary ossification centers

Kronenberg, H. (2003) *Nature*. 423: 332-336.

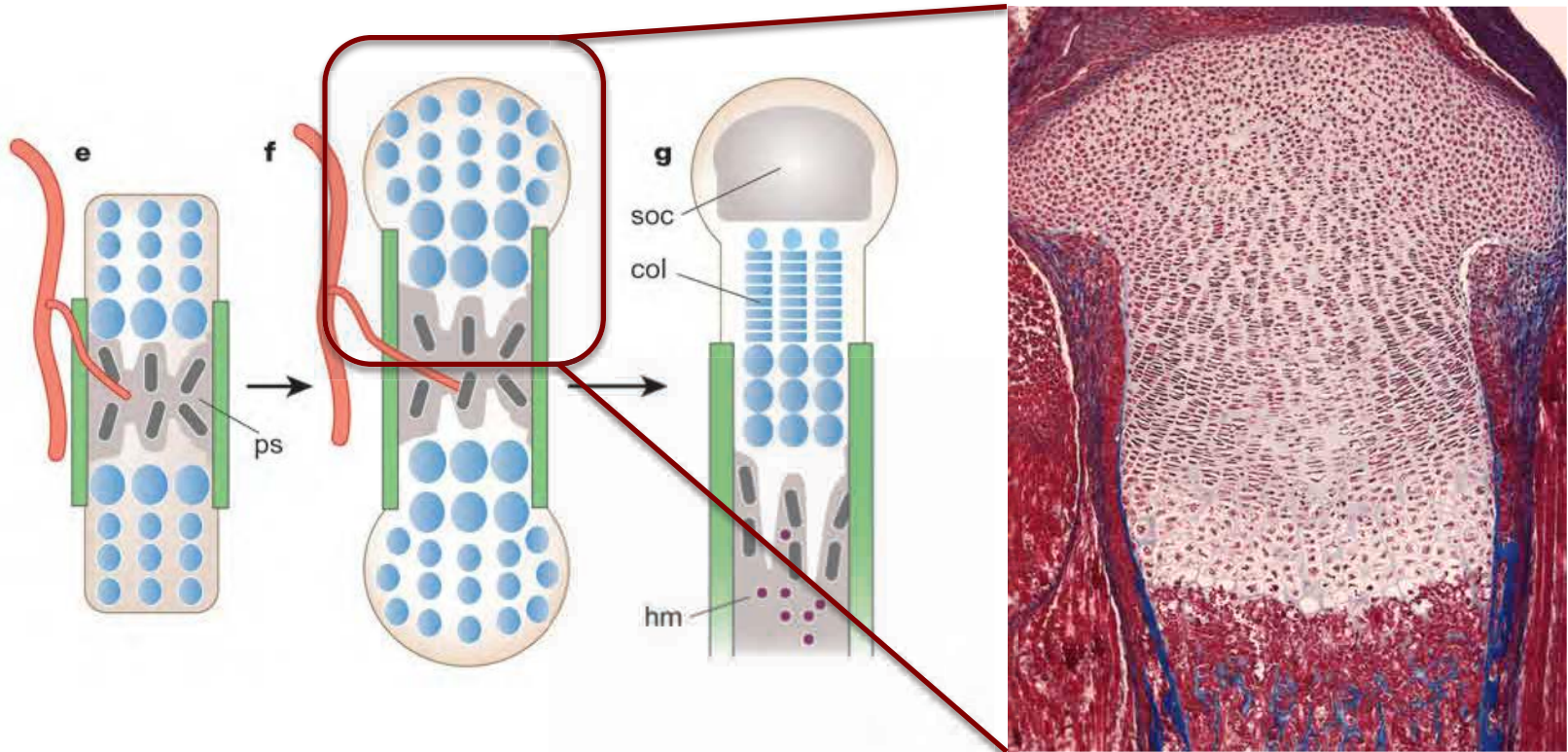
Chondrocyte Maturation – Onset of Hypertrophy



Mouse E14.5 Humerus

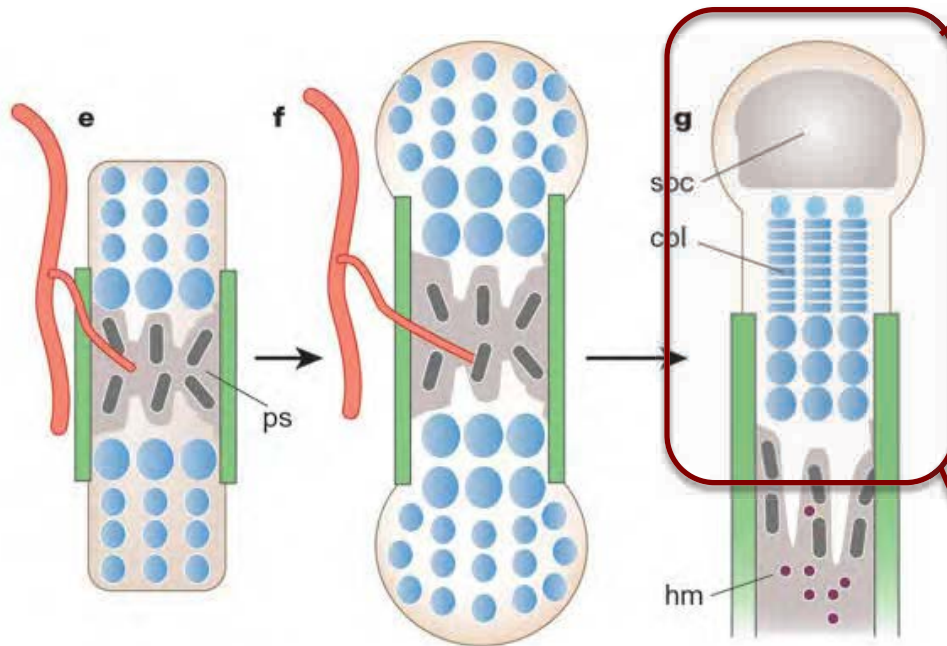
Chondrocyte Maturation – Terminal Hypertrophy

Mouse E18.5 Proximal Tibia

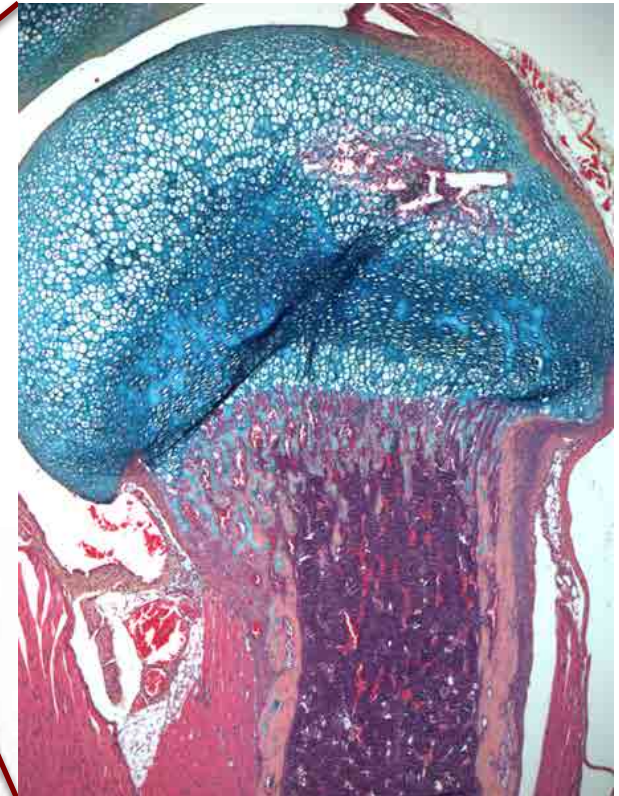


Endochondral Bone Formation

Establishment of Growth Plate and Articular Cartilages

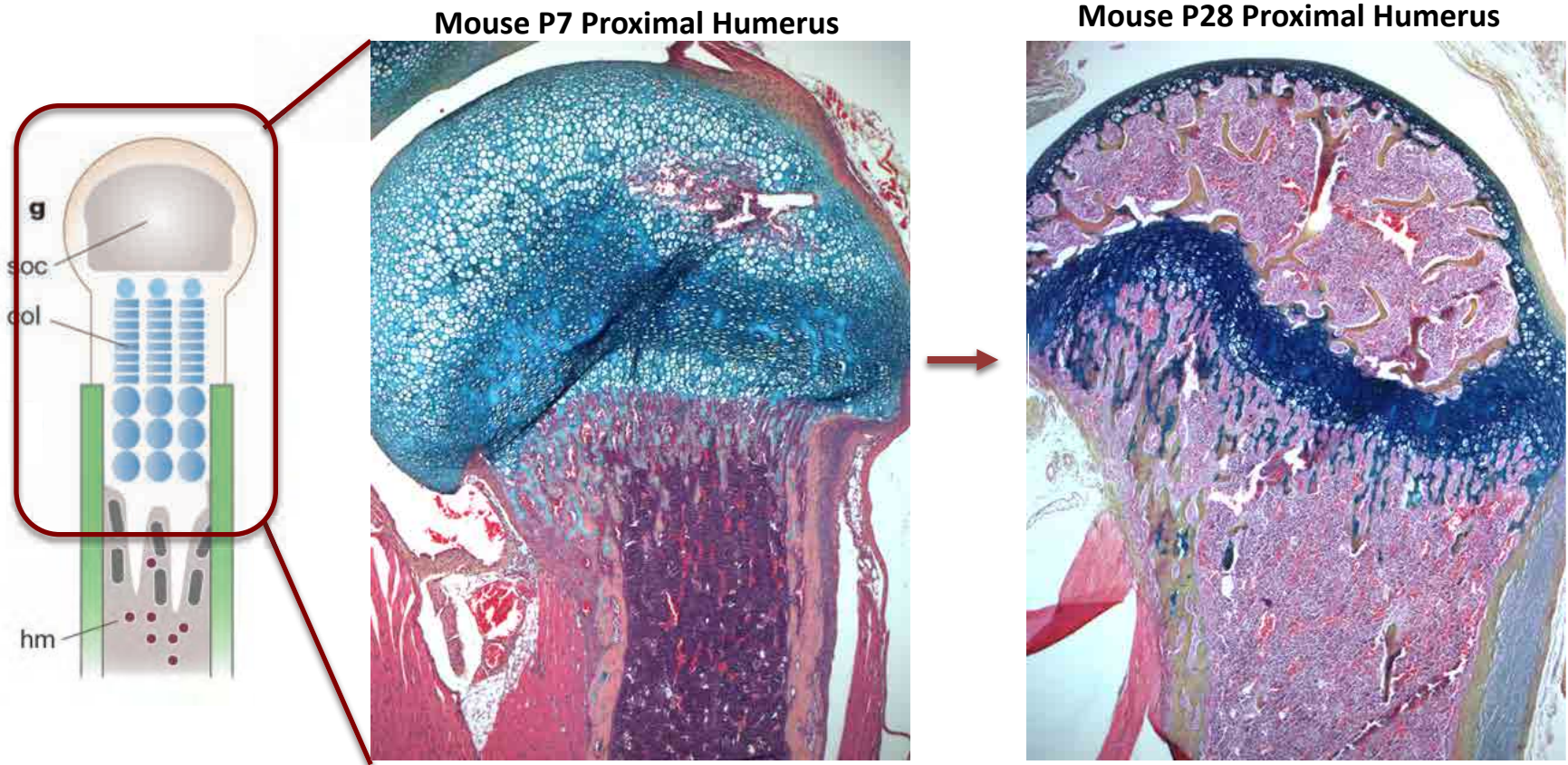


Mouse P7 Proximal Humerus

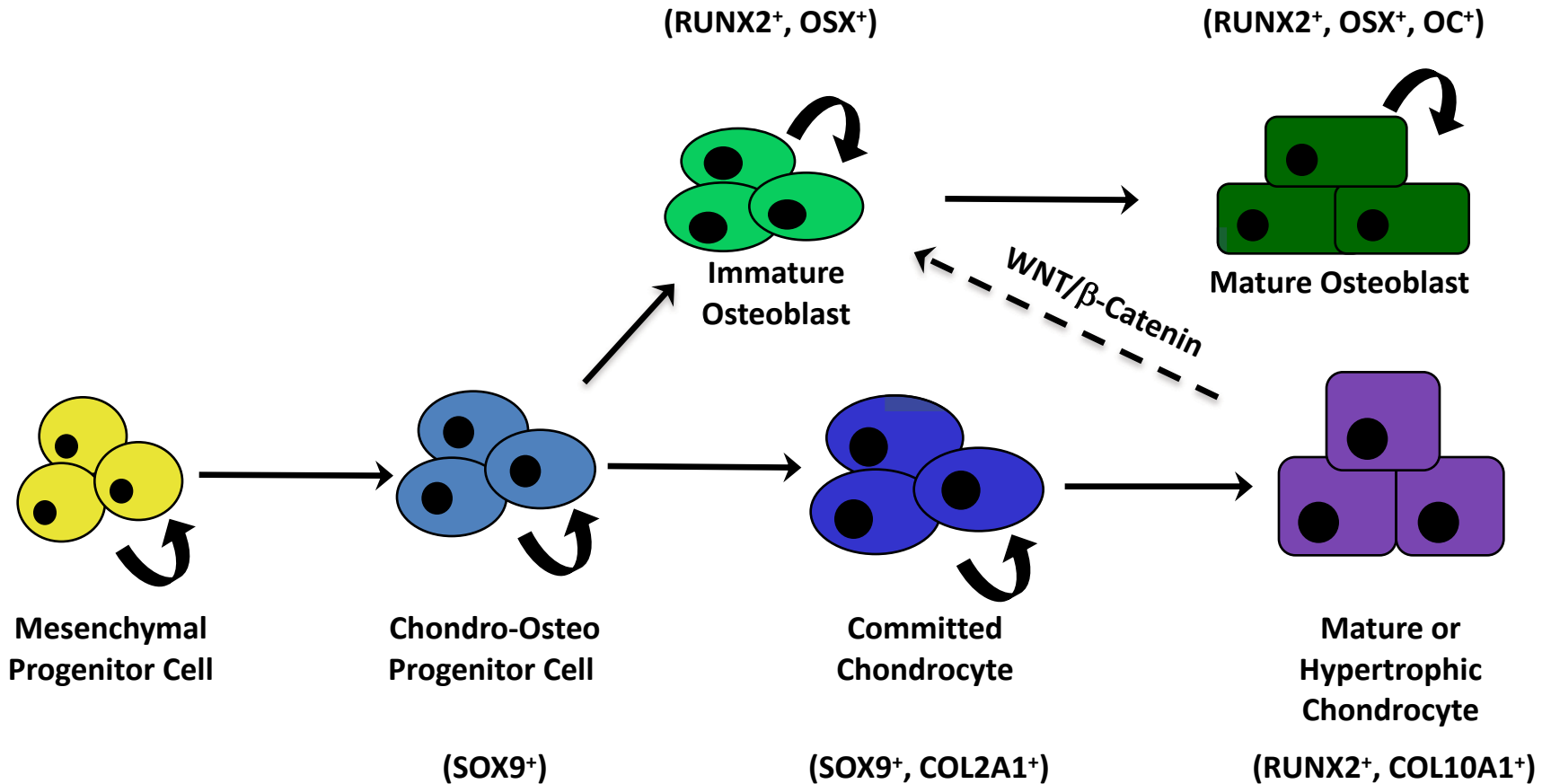


Endochondral Bone Formation

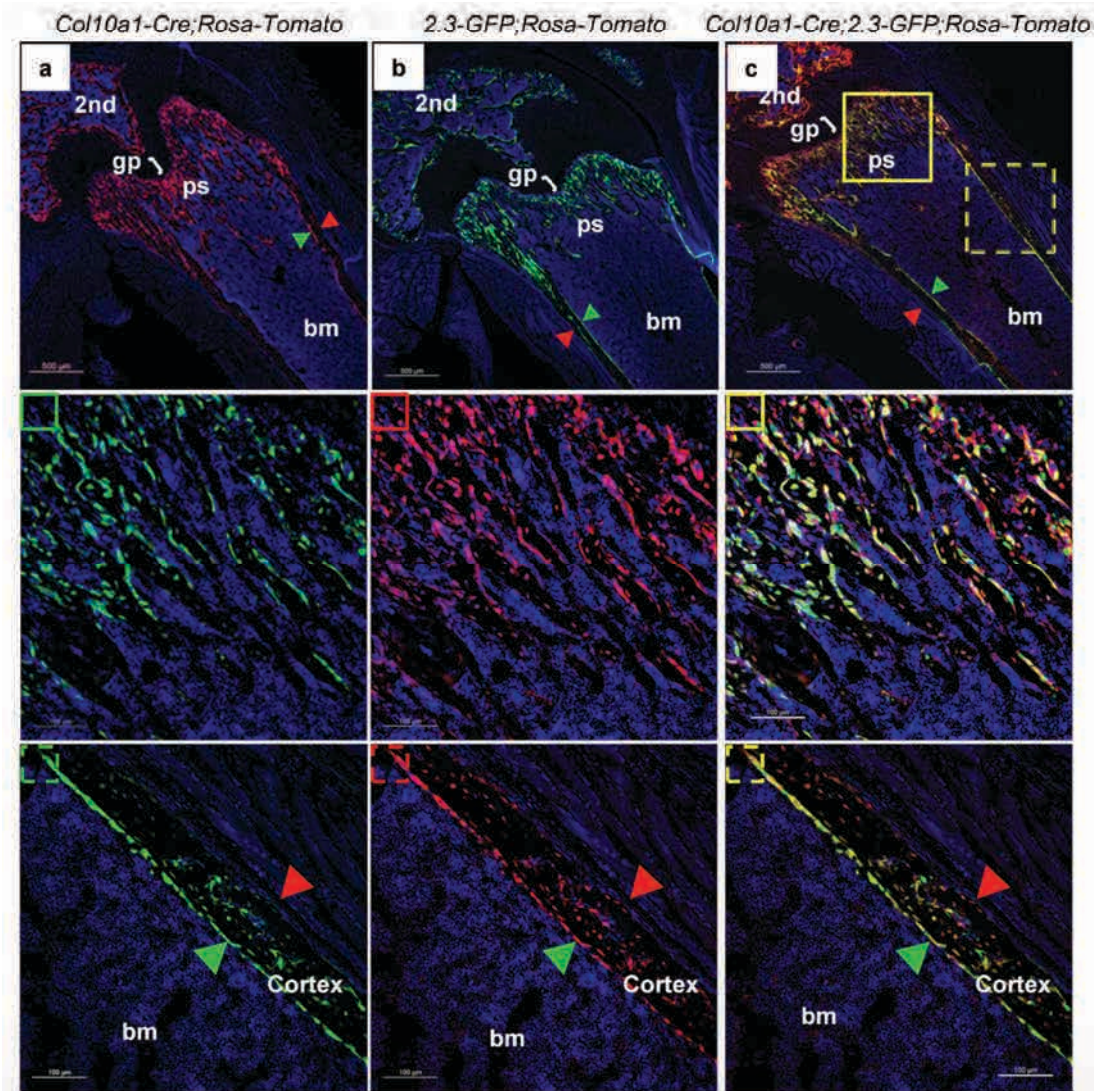
Establishment of Growth Plate and Articular Cartilages



Chondrogenesis and Osteoblastogenesis

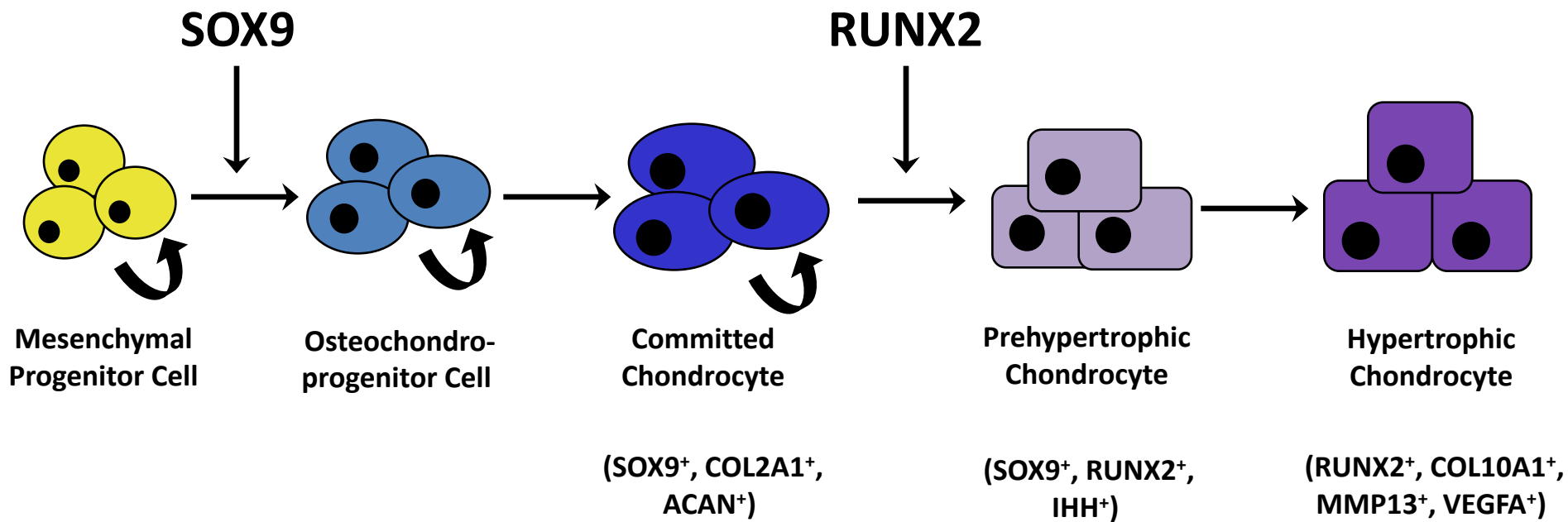


Transdifferentiation of Hypertrophic Chondrocytes to Osteoblasts During Endochondral Bone Formation



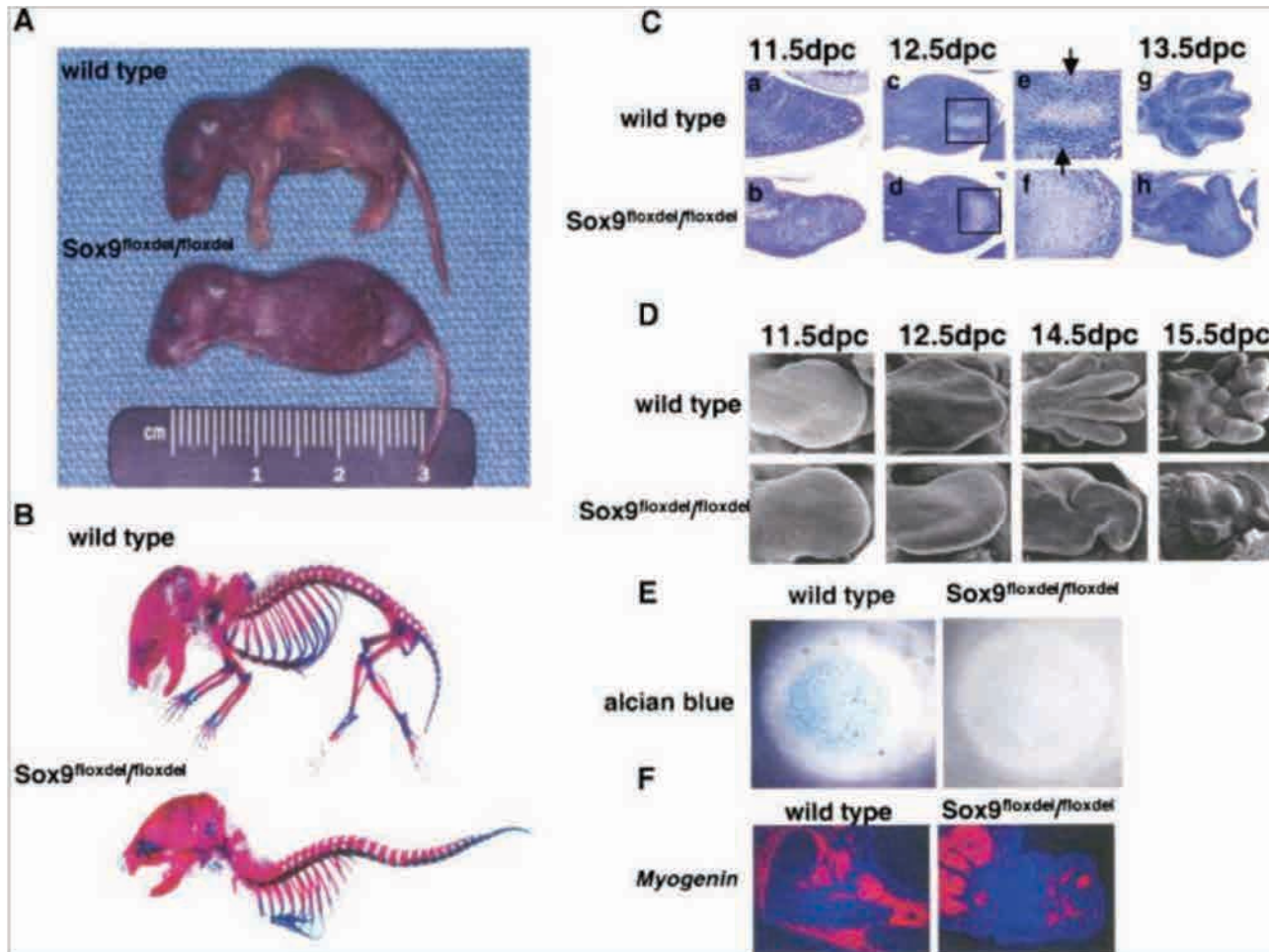
Zhou, et. al. (2014) *Plos Gen.* 12: e1004820.

Molecular Regulation of Chondrocyte Maturation



SOX9 is Required for Chondrogenesis

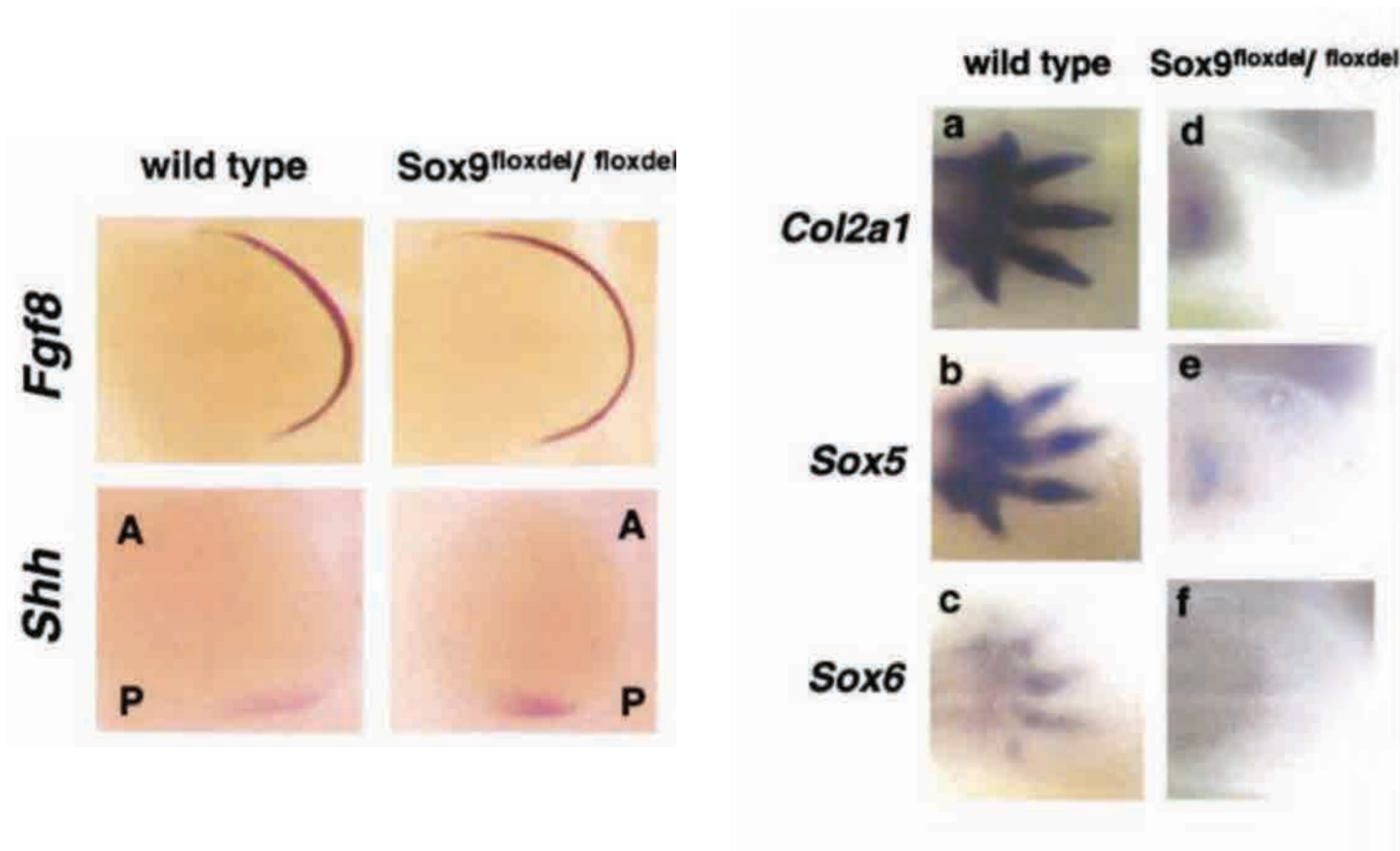
Prx1-Cre (expressed in all MSCs of limb bud approx. E9.5)



Akiyama, et. al. (2002) *Genes Dev.* 16: 2813-2828.

SOX9 is Required for Chondrogenesis

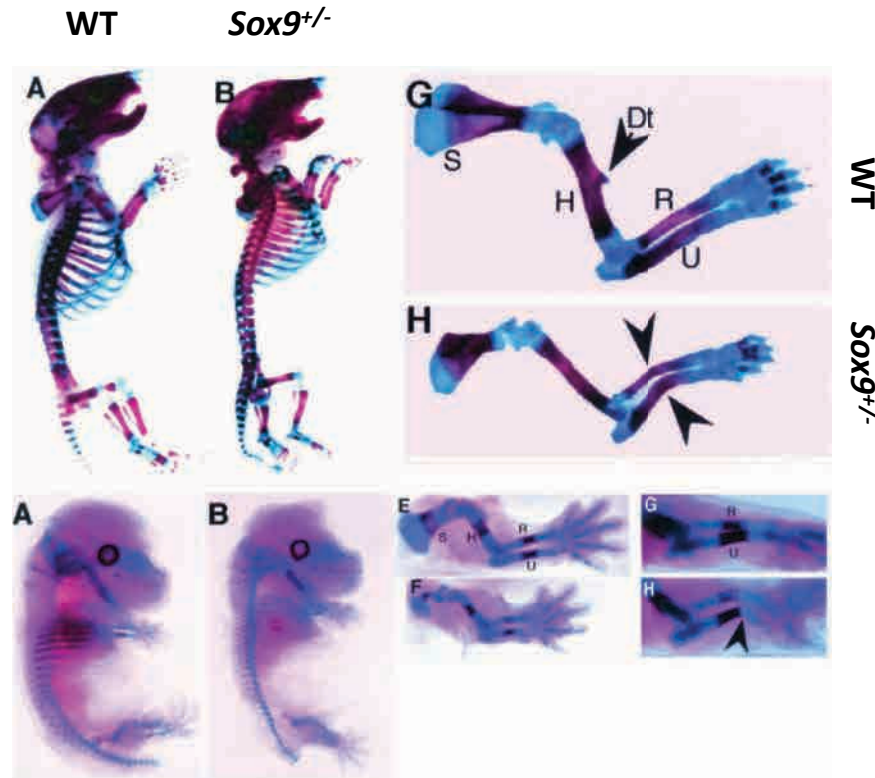
Prx1-Cre (expressed in all MSCs of limb bud approx. E9.5)



Akiyama, et. al. (2002) *Genes Dev.* 16: 2813-2828.

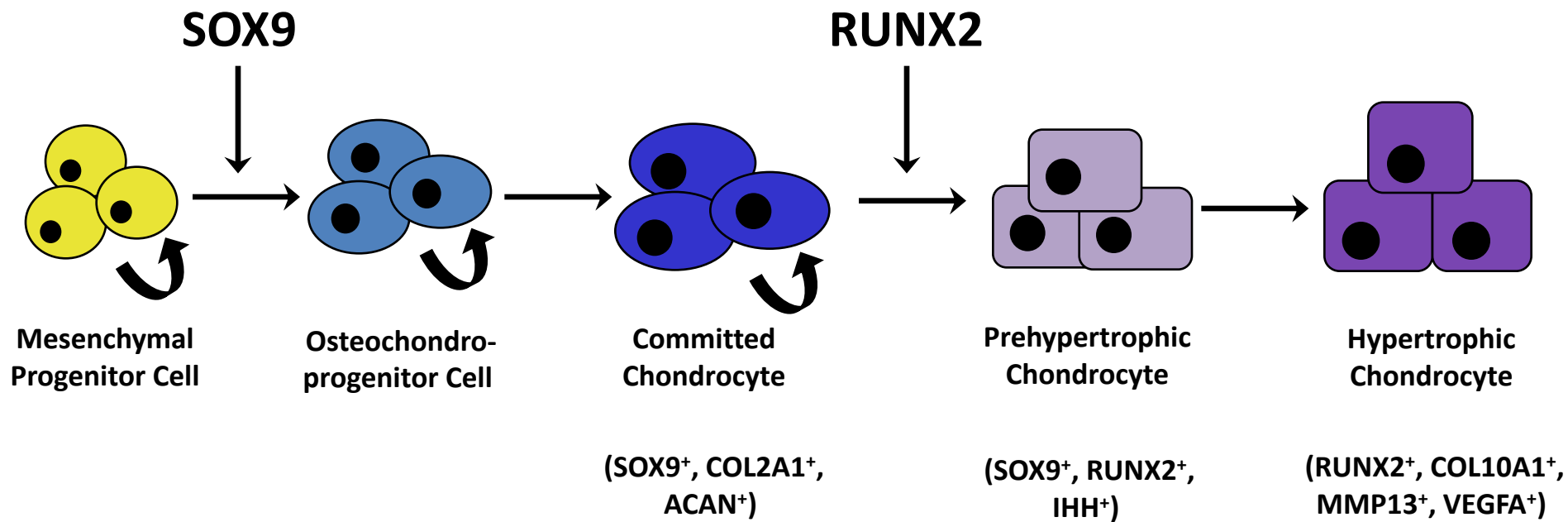
Campomelic Dysplasia

- Autosomal dominant disorder due to haploinsufficiency of *SOX9*
- Clinical features include; bowing of long bones, hypoplasia of the scapula and pelvis, decreased number of ribs, craniofacial abnormalities
- Most affected infants die due to respiratory distress



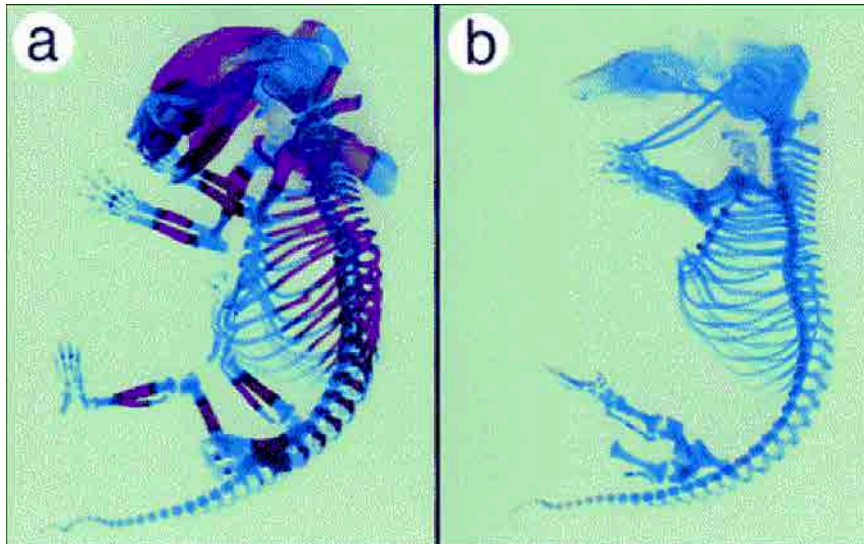
Bi, et. al. (2001) *PNAS*. 98: 6698-6703.

Molecular Regulation of Chondrocyte Maturation



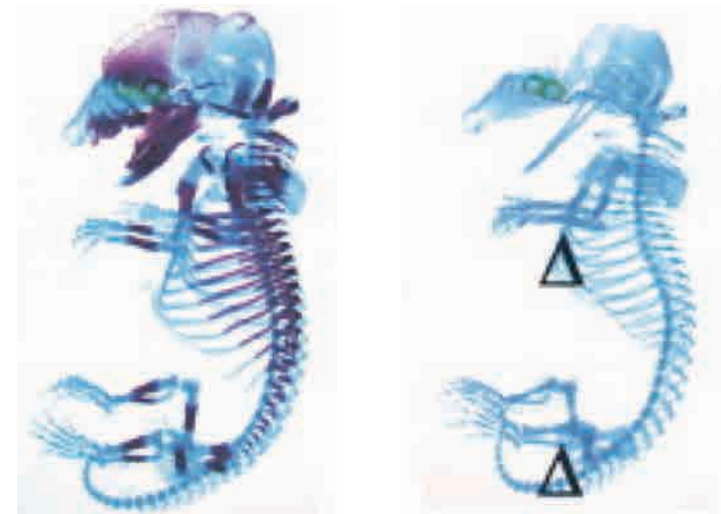
Critical Transcriptional Regulators of Chondrocyte Maturation and Bone Development (Runx2 and Osterix)

Runx2 mutant mice (null)



Otto F et al., (1997) Cell (89)5:765-771.

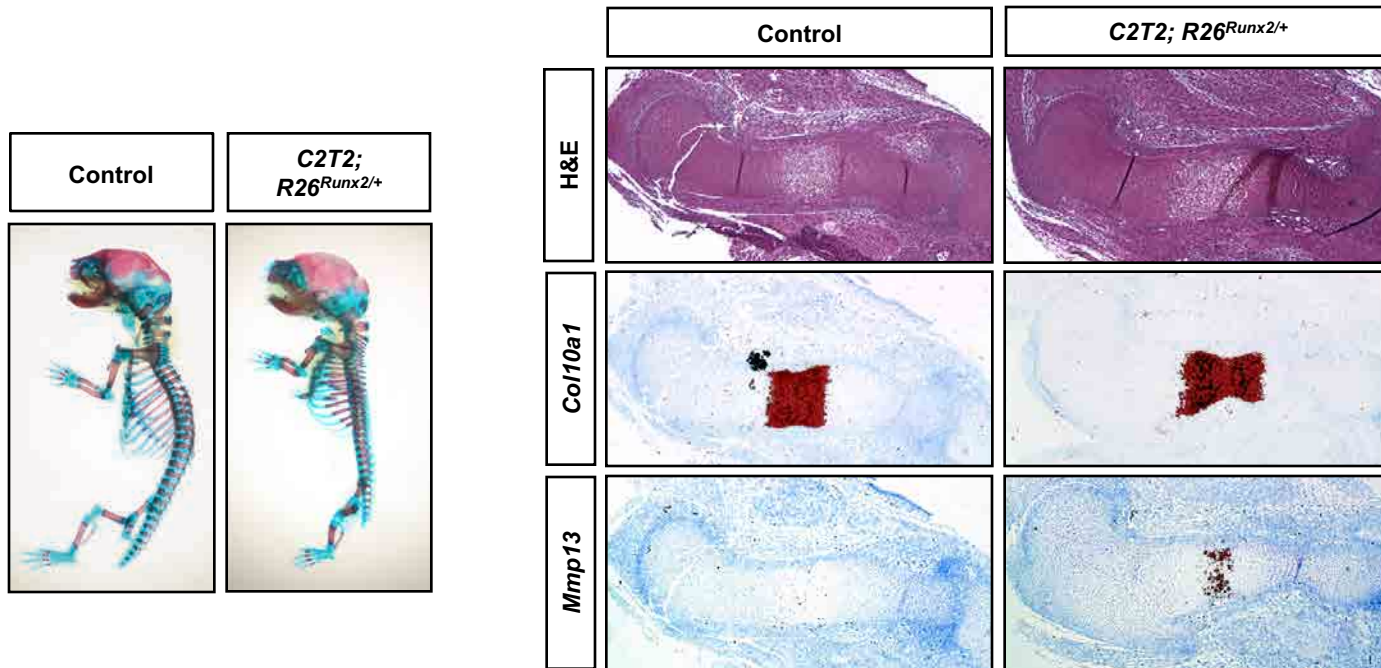
Osterix mutant mice (null)



Nakashima K et al., (2002) Cell (108)1:17-29.

RUNX2 Overexpression Promotes Chondrocyte Maturation

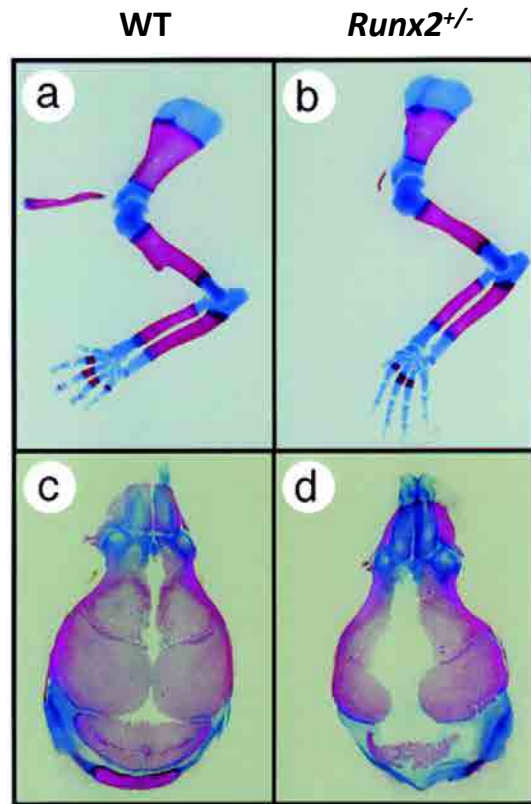
Col2a1-CreER^{T2} (induced at E13.5 for target chondrocytes)



Catheline, et. al. (2019) *JBMR*. doi: 10.1002/jbmr.3737

Cleidocranial Dysplasia (CCD)

- Autosomal dominant disorder due to haploinsufficiency of *RUNX2*
- Clinical features include; hypoplastic clavicles, delayed ossification of the cranial sutures and fontanelles, dental anomalies, and short stature
- At risk for development of osteoporosis and fracture



Otto, et. al. (1997) *Cell*. 89: 765-771.



<http://dentistryandmedicine.blogspot.com/2011/05/cleidocranial-dysostosis-cleidocranial.html>



Supernumerary teeth



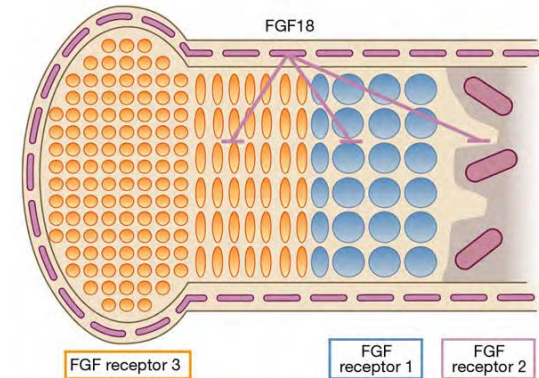
<http://drgstoothpix.com/2014/05/09/case-of-the-week-cleidocranial-dysplasia/>

Human Disorders of Skeletal Patterning, Differentiation, and Growth

- **Polydactyly** (extra digits)
- **Oligodactyly** (missing digits)
- **Amelia** (absence of limbs)
- **Meroamelia** (partial absence of a limb)

- **Brachydactyly** (shortened digits)
- **Campomelic Dysplasia** (*SOX9* haploinsufficiency)
- **Ellis-van Creveld Syndrome** (*EVC* mutations)
- **Cleidocranial Dysplasia** (*RUNX2* haploinsufficiency)

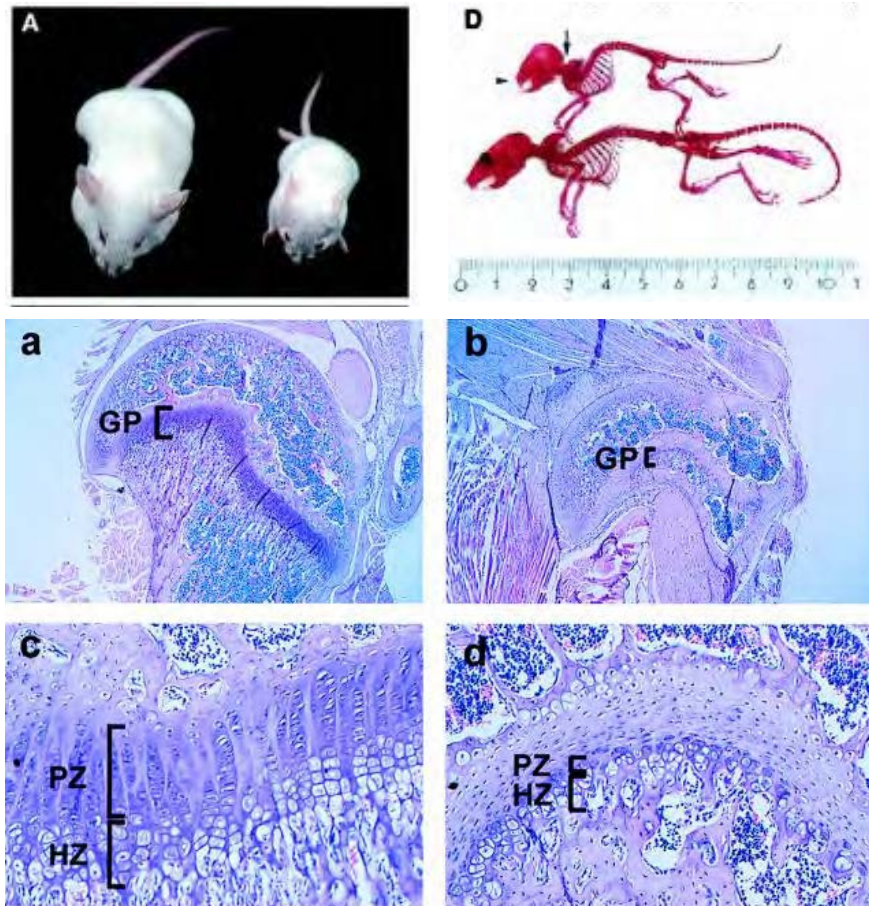
- **Dwarfing Chondrodysplasias** (caused by activating *FGFR3* mutations)
 - **Hypochondroplasia** (least severe; short stature)
 - **Achondroplasia** (most common; short stature, frontal bossing)
 - **Thanatophoric Dysplasia** (most severe; most patients die of respiratory distress following birth)



For review, Kornak and Mundlos (2003) *Am. J. Hum. Genet.* 73: 447-474.

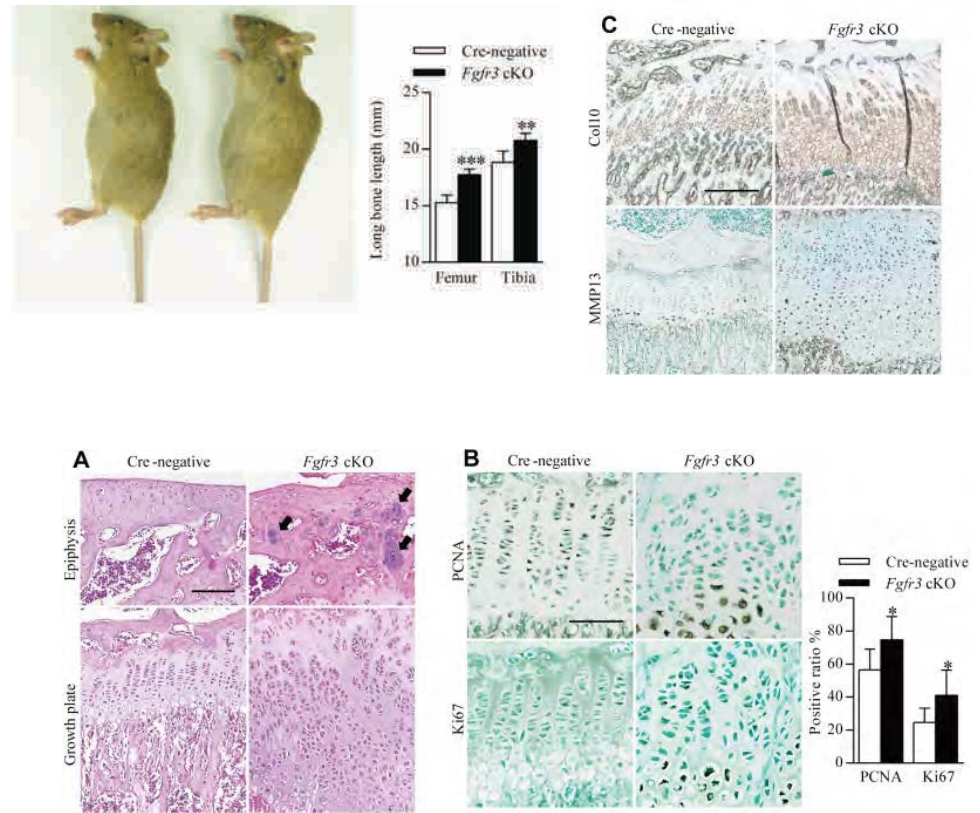
FGFR3 Mouse Models

FGFR3 Activation (FGFR3^{G374Rneo-/+}) Achondroplasia model



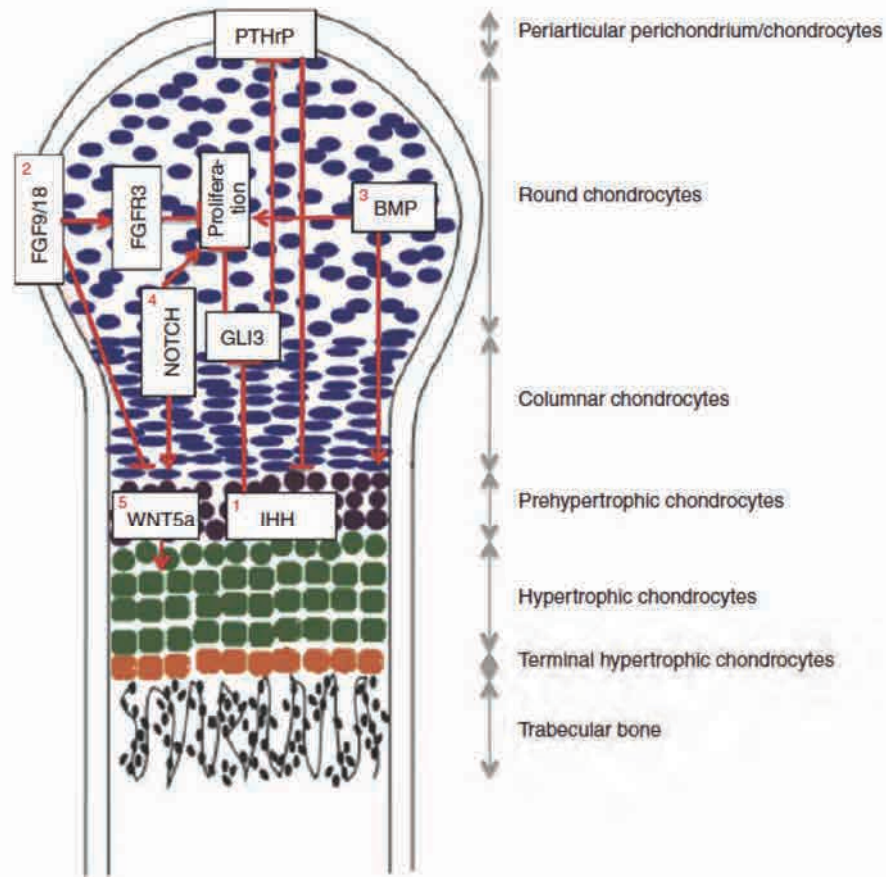
Wang, et. al. (1999) PNAS. 96: 4455-4460.

Chondrocyte-specific loss of *Fgfr3* leads to enhanced proliferation and maturation



Zhou, et. al. (2015) Plos Genet. 11: ePub.

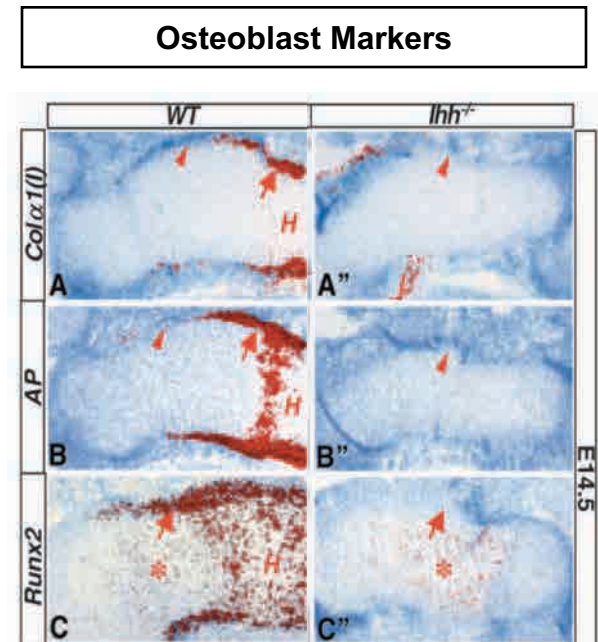
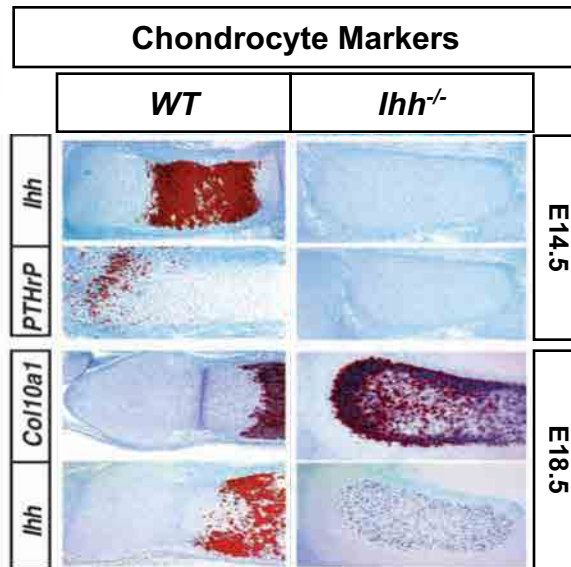
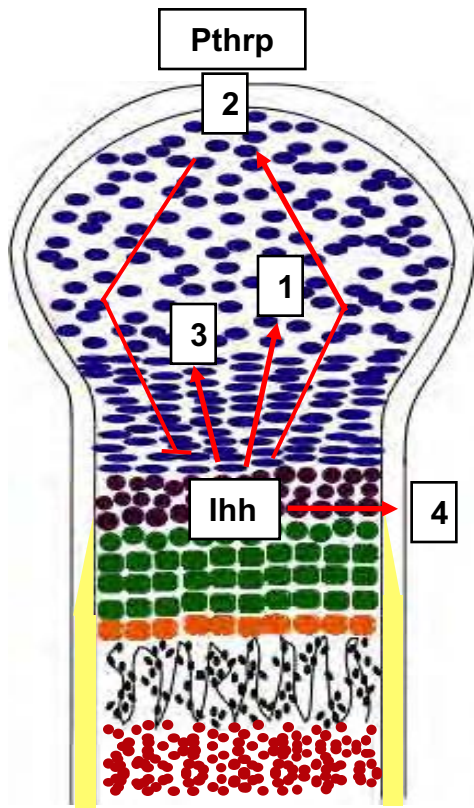
Signaling Pathways that Regulate Growth Plate Development



Long & Ornitz (2013) *CSH Perspect Biol.* 5:a008334.

Ihh/Pthrp Signaling During Cartilage and Bone Development

- Ihh signaling induces chondrocyte proliferation, delays chondrocyte differentiation, and is required for early osteoblast maturation.

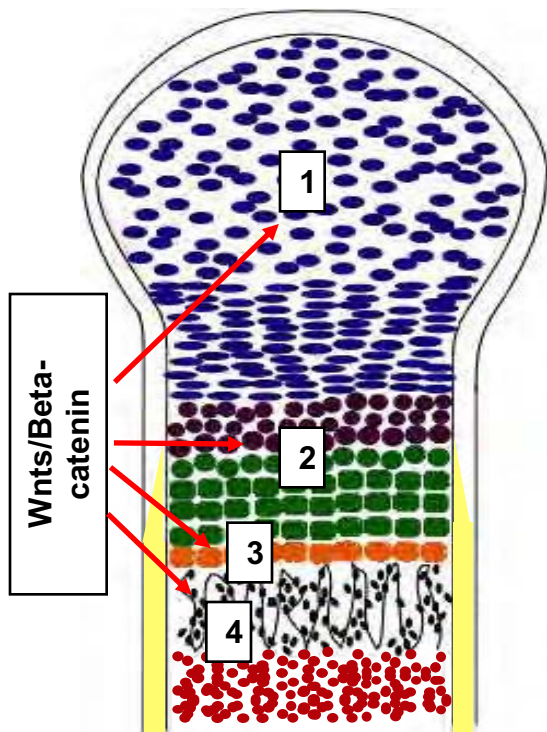


Long, et al. (2001) *Development*. 128:5099-5108.

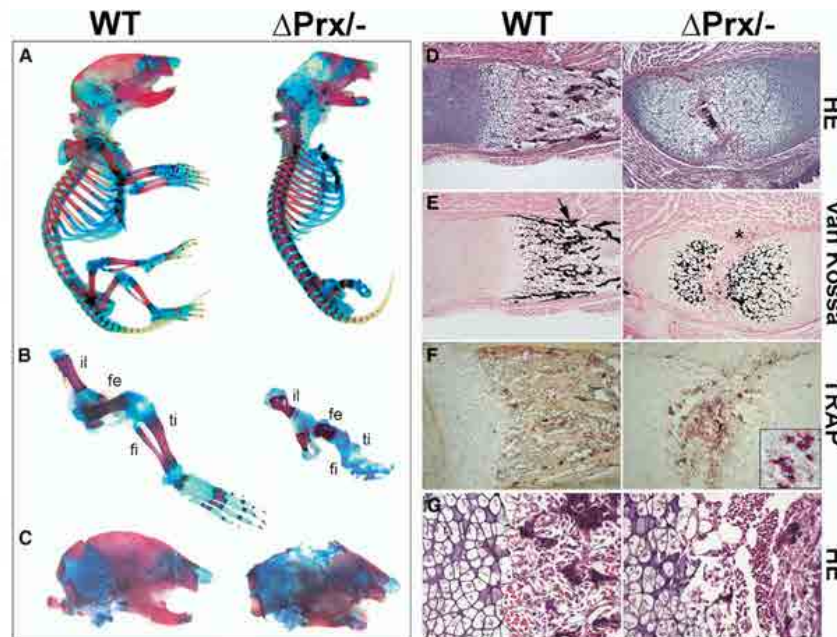
Hu, et. al. (2005) *Development*. 132:49-60.

Wnt/ β -Catenin Signaling During Cartilage and Bone Development

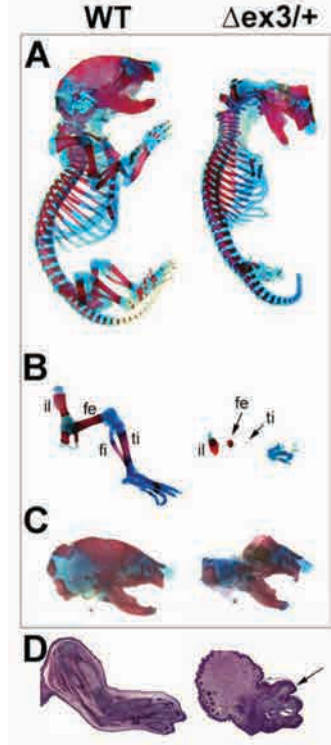
- Wnt/ β -catenin signaling induces chondrocyte proliferation, differentiation, and osteoblast maturation in committed cells.
- Wnt/ β -catenin signaling suppresses chondrogenic and osteogenic differentiation in mesenchymal progenitor cells.



β -catenin Loss-of-function (Prx1-Cre)

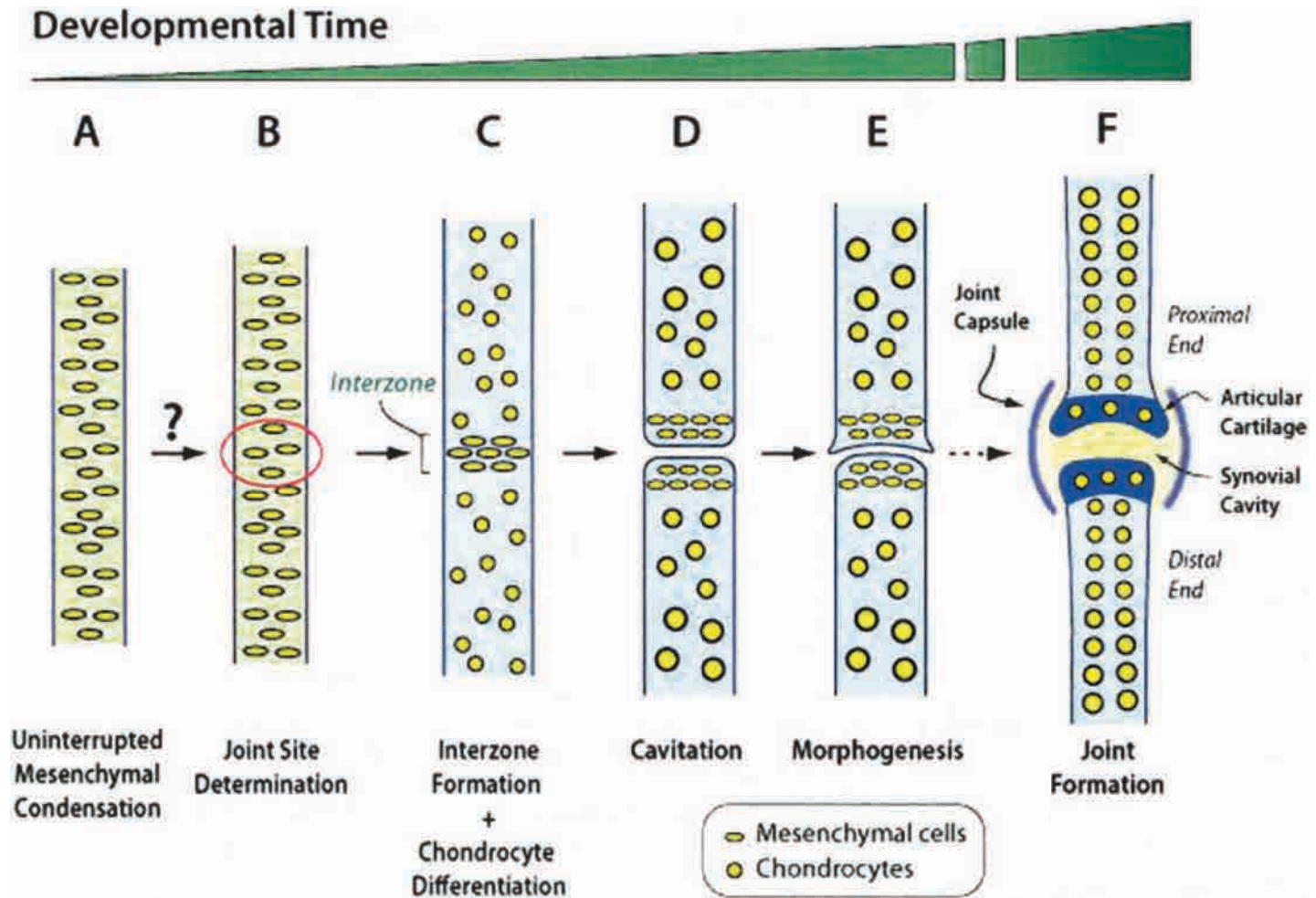


Gain-of-function



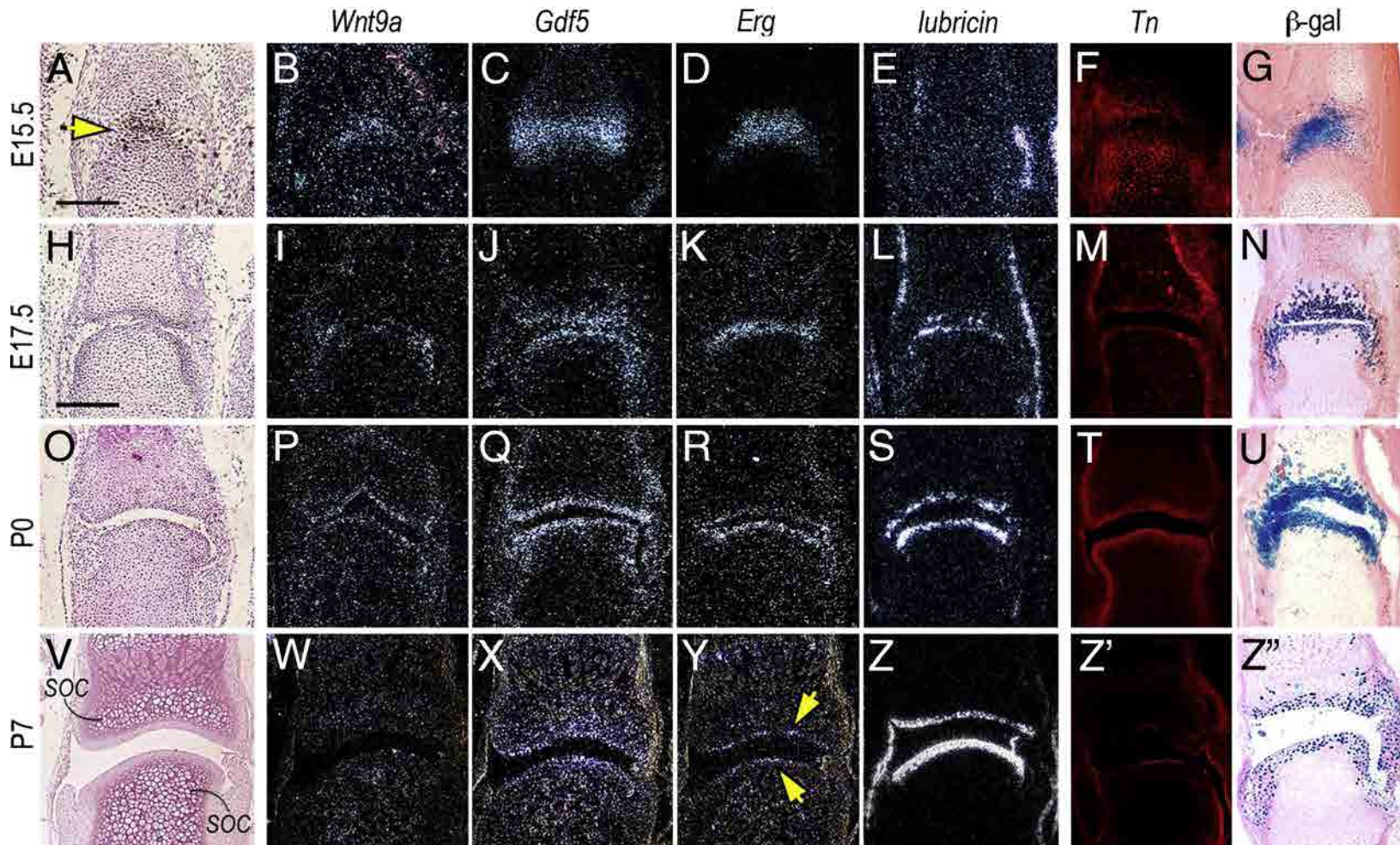
Hill, et al. (2005) *Developmental Cell*. 8:727-738.

Synovial Joint Development During Embryogenesis



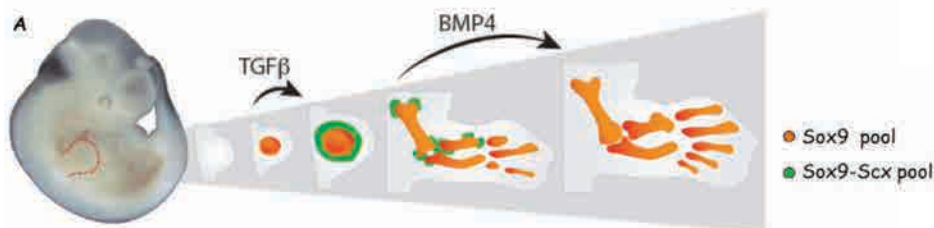
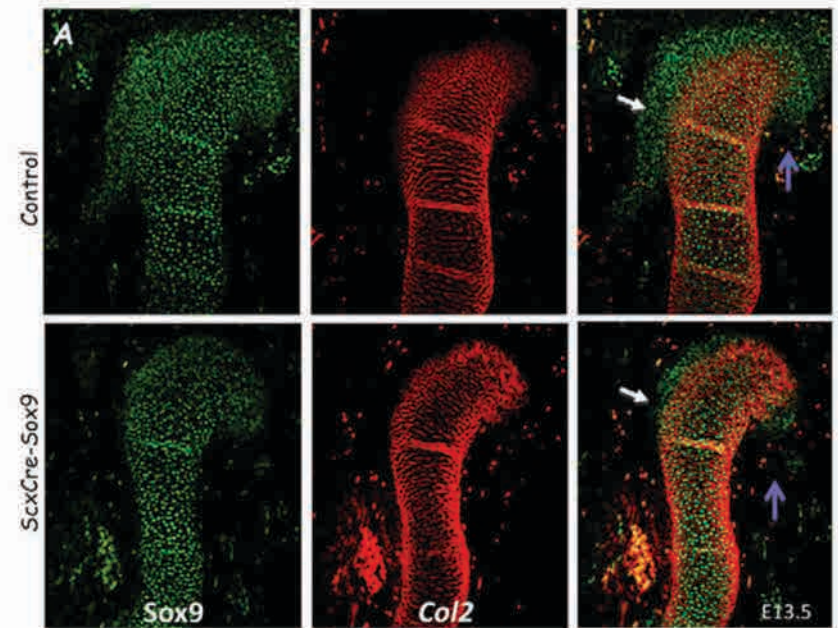
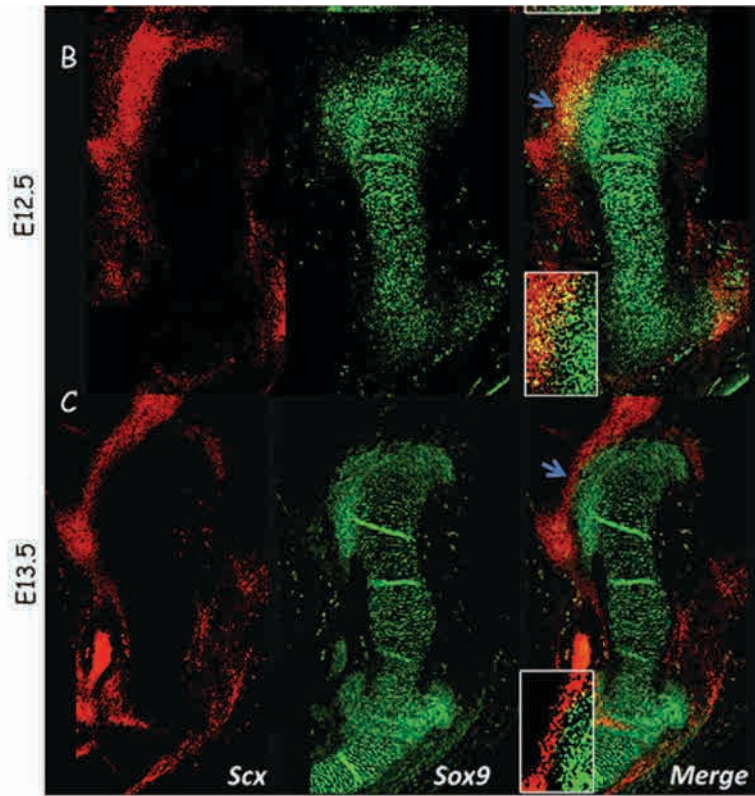
Pacifici, et al. (2005) Birth Defects Res. 75:237-248.

Molecular Regulation of Joint Development



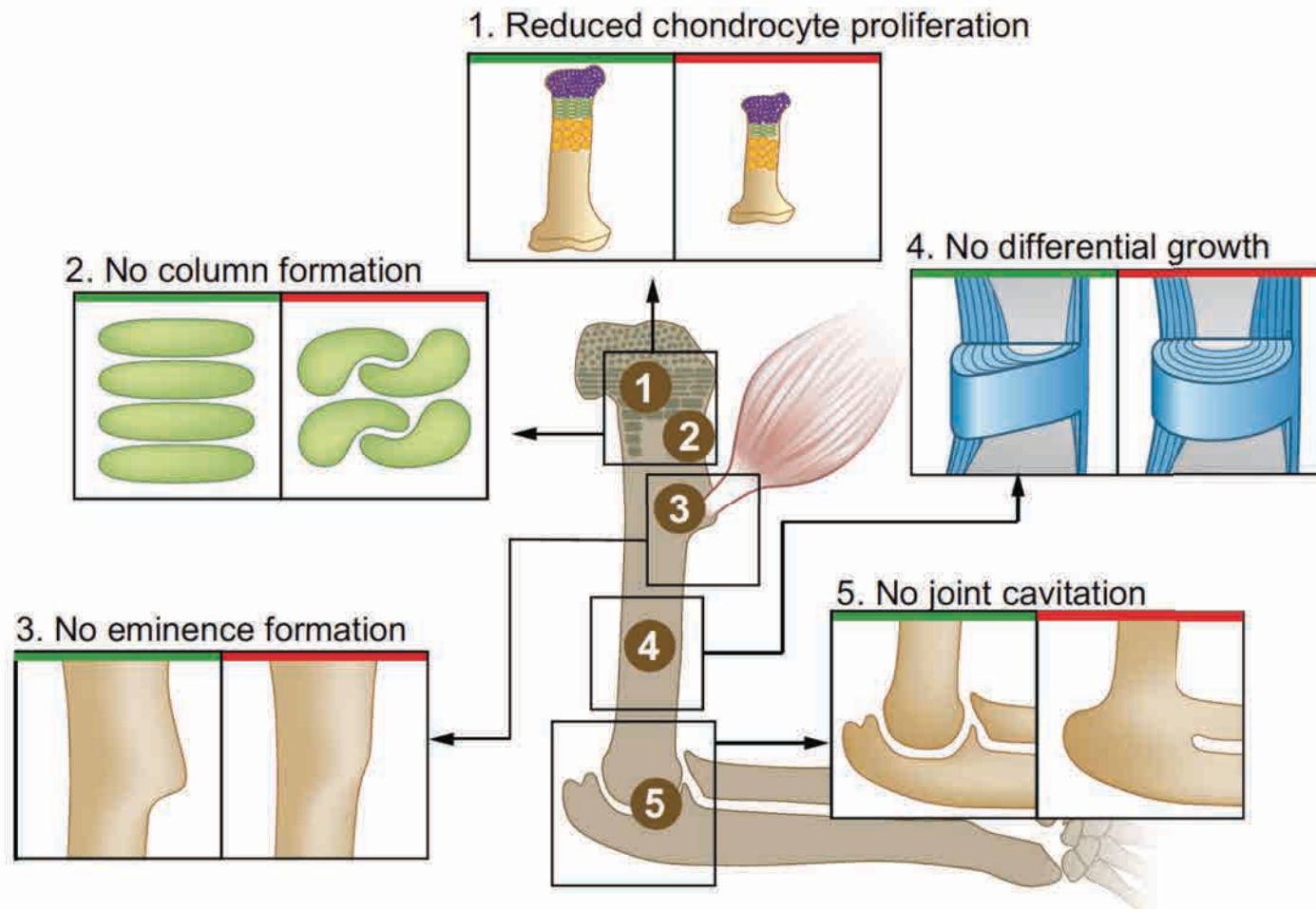
Koyama, et al. (2008) Developmental Biology. 1:62-73.

Bone Eminence Development Requires Sox9 Expression



Blitz, et al. (2013) *Development*. 140:2680-90.

Reduced Mechanical Forces Affect Musculoskeletal Development



Felsenthal, et al. (2017) Development. 144: 4271-83.